

TOPICAL REVIEW

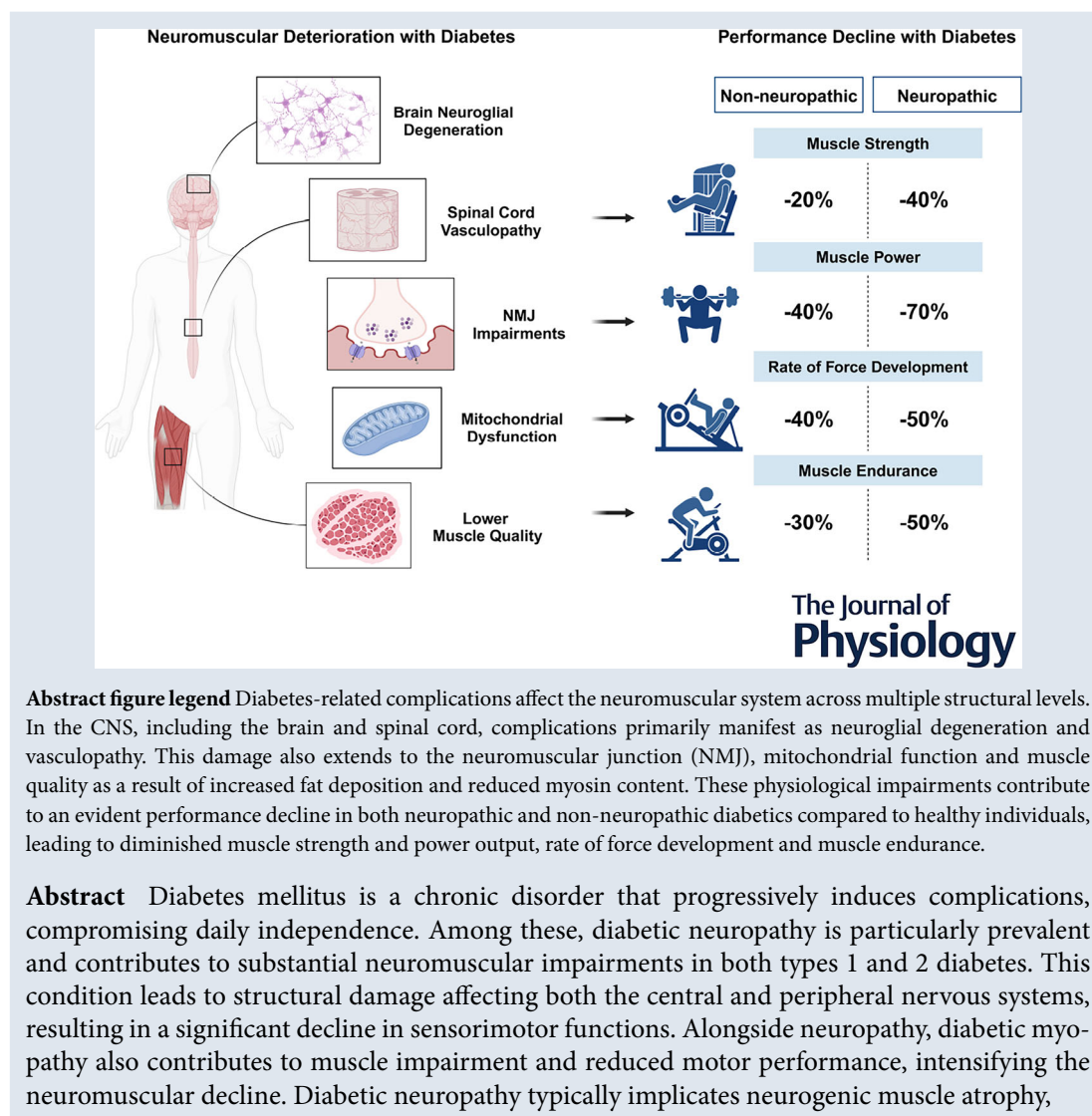
Physiological mechanisms of neuromuscular impairment in diabetes-related complications: Can physical exercise help prevent it?

Edoardo Lecce , Alessio Bellini , Giuseppe Greco , Fiorella Martire , Alessandro Scotto di Palumbo, Massimo Sacchetti  and Ilenia Bazzucchi 

Laboratory of Exercise Physiology, Department of Movement, Human, and Health Sciences, University of 'Foro Italico', Rome, Italy

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Abstract figure legend Diabetes-related complications affect the neuromuscular system across multiple structural levels. In the CNS, including the brain and spinal cord, complications primarily manifest as neuroglial degeneration and vasculopathy. This damage also extends to the neuromuscular junction (NMJ), mitochondrial function and muscle quality as a result of increased fat deposition and reduced myosin content. These physiological impairments contribute to an evident performance decline in both neuropathic and non-neuropathic diabetics compared to healthy individuals, leading to diminished muscle strength and power output, rate of force development and muscle endurance.

Abstract Diabetes mellitus is a chronic disorder that progressively induces complications, compromising daily independence. Among these, diabetic neuropathy is particularly prevalent and contributes to substantial neuromuscular impairments in both types 1 and 2 diabetes. This condition leads to structural damage affecting both the central and peripheral nervous systems, resulting in a significant decline in sensorimotor functions. Alongside neuropathy, diabetic myopathy also contributes to muscle impairment and reduced motor performance, intensifying the neuromuscular decline. Diabetic neuropathy typically implicates neurogenic muscle atrophy,

motoneuron loss and clustering of muscle fibres as a result of aberrant denervation-reinnervation processes. These complications are associated with compromised neuromuscular junctions, where alterations occur in pre-synaptic vesicles, mitochondrial content and post-synaptic signalling. Neural damage is intensified by chronic hyperglycaemia and oxidative stress, exacerbating vascular dysfunction and reducing oxygen delivery. These complications imply a severe decline in neuromuscular performance, evidenced by reductions in maximal force and power output, rate of force development and muscle endurance. Furthermore, diabetes-related complications are compounded by age-related degenerative changes in long-term patients. Aerobic and resistance training offer promising approaches for managing blood glucose levels and neuromuscular function. Aerobic exercise promotes mitochondrial biogenesis and angiogenesis, supporting metabolic and cardiovascular health. Resistance training primarily enhances neural plasticity, muscle strength and hypertrophy, which are crucial factors for mitigating sarcopenia and preserving functional independence. This topical review examines current evidence on the physiological mechanisms underlying diabetic neuropathy and the potential impact of physical activity in counteracting this decline.

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Corresponding author I. Bazzucchi: Department of Movement, Human and Health Sciences, Laboratory of Exercise Physiology, University of 'Foro Italico', Piazza Lauro De Bosis, 6, Rome 00135, Italy. Email: ilenia.bazzucchi@uniroma4.it

Introduction

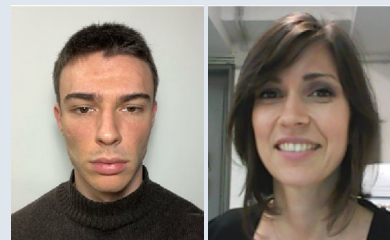
Diabetes mellitus is a chronic metabolic disorder affecting millions worldwide, and it is associated with various complications (Harding et al., 2019; Ong et al., 2023; Shaw et al., 2010). Diabetic neuropathy (DN) is among the most common complications, progressively impairing both the central and peripheral nervous systems in type 1 (T1D) and type 2 diabetes (T2D) (Feldman et al., 2019). DN primarily affects somatic and autonomic nerves in a distal, symmetrical pattern, progressing according to fibre length (Forbes & Cooper, 2013; Said, 2007; Singh et al., 2014), ultimately resulting in impaired neural function, sarcopenia and neurogenic muscle fibre atrophy, implicating neuromuscular deconditioning (Allen et al., 2016; Haines et al., 2022; Wong et al., 2017).

Over time, both T1D and T2D are also linked to diabetic myopathy, a disorder affecting muscle mass and function as a result of chronic hyperglycaemia,

altered muscle metabolism and reduced skeletal muscle regenerative capacity (D'Souza et al., 2013; Hernández & Camilo Vanegas, 2015; Monaco et al., 2019). Together with neuropathic complications, these factors contribute to the neuromuscular function decline associated with diabetes. In addition, long-term T2D is typically accompanied by older age (Assar et al., 2019), intensifying degenerative changes through the combined effects of ageing and diabetes (Hepple & Rice, 2016; Saeedi et al., 2019; Xu et al., 2018). This progressive neuromuscular deterioration notably impacts the quality of life and daily independence (Parasoglou et al., 2017; Wong et al., 2017), together with a marked decline in physical performance (Luo et al., 2022; Orlando et al., 2022).

In response to the multifactorial nature of neuromuscular deterioration in diabetes, there is growing interest in examining its physiological determinants and associated complications (Allen et al., 2016; Feldman et al., 2019; Le Corre et al., 2023; Orlando et al.,

Edoardo Lecce is a PhD candidate at the University of Rome 'Foro Italico', where he specialised in electromyographic research on human motor units during his Master's degree. His work includes non-invasive high-spatial-resolution electromyography to characterise motor unit activity across different populations. His research explores adaptations to resistance training and mechanical perturbations, aiming to expand the understanding of neuromuscular function and its practical applications in healthy individuals and those affected by chronic disorders. **Ilenia Bazzucchi** has a PhD in Sport Science and Health, currently Associate Professor in Human Physiology and Head of the Exercise Physiology Laboratory at the University of Rome 'Foro Italico'. She has conducted research since 1998 at the Human Physiology Institute of the University of Rome 'Sapienza' and, in 2002, she was a visiting researcher at the Institute for Biomedical Science at the Strathclyde University of Glasgow (UK). Her research work mainly concerns the study of neuromuscular control by means of surface electromyography, neuromuscular effects of exercise and nutritional interventions in athletes, as well as healthy and pathological individuals.



2022; Singh et al., 2014). Although prior reviews have addressed individual aspects of diabetic neuropathy, a comprehensive analysis encompassing central and peripheral mechanisms, alongside the potential of physical exercise to mitigate these adverse effects, remains limited. This topical review aims to synthesise current knowledge on structural and functional alterations associated with diabetic neuropathy in the advanced condition stage, offering an integrated perspective on its pathophysiology and the role of exercise in alleviating diabetes-related complications.

The literature search was conducted using the following terms on PubMed/MEDLINE/MeSH and Scopus: ('diabetes mellitus' OR 'diabetes' OR 'diabetic neuropathy' OR 'diabetes-related complications' OR 'diabetic complications') AND ('ageing' OR 'aging' OR 'older adults') AND ('neuromuscular' OR 'motor unit' OR 'muscle impairment' OR 'muscle weakness' OR 'sarcopenia' OR 'neuromuscular junction') AND ('exercise' OR 'physical activity' OR 'exercise intervention' OR 'resistance training' OR 'endurance training' OR 'aerobic exercise' OR 'combined exercise'). Additional articles suggested or related to these search results were also reviewed and included where relevant.

Differential pathogeneses in T1D and T2D

T1D typically manifests early in life, often during childhood or adolescence (peaking at ~10–14 years of age), and is characterised by an acute and rapid onset of hyperglycaemia following autoimmune destruction of insulin-producing β -cells in the pancreas (Sempere-Bigorra et al., 2021). This autoimmune response is driven by genetic susceptibility and environmental factors, such as viral infections, implicating an absolute insulin deficiency (Ozougwu, 2013). Because of this sudden onset, individuals with T1D experience swift blood glucose fluctuations, underscoring consistent glycaemic control to mitigate the risk of complications (Cernea & Raz, 2021).

T2D develops gradually and is typically diagnosed in adulthood (peaking at ~55–59 years of age), although rising obesity rates and sedentary lifestyles have led to increased incidence among younger populations (Sempere-Bigorra et al., 2021). T2D pathogenesis primarily depends on insulin resistance, where body tissues become less responsive to insulin. This condition usually initiates with prolonged prediabetes and hyperinsulinaemia as the pancreas compensates for insulin resistance. Over time, β -cell dysfunction ensues, resulting in a relative insulin deficiency. The slow progression of T2D, shaped by genetic predispositions, lifestyle factors and ageing, often leads to established neuropathic complications by the time of diagnosis (Ozougwu, 2013).

Neuropathy development in T1D and T2D. Although diabetic neuropathy is a prevalent complication in both types of diabetes, evidence highlights distinct pathogenetic mechanisms underlying this condition in T1D and T2D (Sempere-Bigorra et al., 2021; Zhu et al., 2024). The lifetime prevalence of diabetic neuropathy is high, affecting up to 30% of individuals with T1D and 50% of those with T2D, although the timing of onset varies between these groups. Neuropathy symptoms in T1D typically appear after a prolonged period (~14–20 years post-diagnosis), with peak prevalence observed after 30 years of age, whereas, in T2D, symptoms emerge after a shorter duration (5–10 years post-diagnosis), peaking after 55 years of age (Feldman et al., 2019; Hicks & Selvin, 2019; Pop-Busui et al., 2017; Sempere-Bigorra et al., 2021).

The pathogenesis of T1D-related neuropathy is associated with disruptions in lipid biosynthesis and cholesterol metabolism, whereas, in T2D, it is linked to aberrant activity of nuclear factor-kappa B, peroxisome proliferator-activated receptor γ coactivator-1 α (PGC1 α) and mitogen-activated protein-kinase pathways involved in insulin signalling (Hur et al., 2016; Ng et al., 2024). Furthermore, studies analysing diabetes-related genes and pathways in the sciatic nerve of diabetic mice have identified distinct patterns of gene expression. In T1D, differentially expressed genes are primarily located in the nucleoplasm, affecting transcriptional regulation, whereas, in T2D, they are predominantly at cellular junctions and impact ion transport processes (Gu et al., 2018).

T1D and T2D present alterations in the neural system and the skeletal muscle (Fig. 1), contributing to a severe decline in motor performance, discussed in the next section.

Diabetic-related complications of the neural system

Individuals affected by diabetic neuropathy experience complications that extend beyond the peripheral nerves, impacting the spinal cord and supraspinal areas (Selvarajah et al., 2006; Ved et al., 2018). This widespread impairment disrupts both motor and somatosensory functions, contributing to motor deficits and neuropathic pain (Feldman et al., 2017; Leitzelar & Koltyn, 2021; Meacham et al., 2017; Woolf, 2011; Zhang et al., 2021). Numerous mechanisms contribute to the neuromuscular impairments observed in both neuropathic and non-neuropathic diabetic individuals.

Anatomical determinants. Diabetic neuropathy primarily targets sensory nerves (Ramji et al., 2007; Zochodne et al., 2001), with motoneurons being affected at a later stage (Feldman et al., 2019; Forbes & Cooper, 2013). This progression is attributed to the anatomical

positioning of sensory neurons in the dorsal root ganglia, which lie outside the blood–brain barrier, becoming highly vulnerable to diabetes-related oxidative stress. By contrast, the cell bodies of spinal motoneurons reside within the ventral horn and are somewhat protected (Ballabh et al., 2004; Feldman et al., 2017). Additionally, neuropathic impairments are more pronounced in the lower limbs, probably as a result of the combined effects of lower glucose clearance compared to the upper extremities and the longer nerve fibre length (Forbes & Cooper, 2013; Olsen et al., 2005; Sacchetti et al., 2013).

Diabetic neuropathy is characterised by vascular deterioration, which includes a high incidence of atherosclerosis, capillary basement membrane thickening and endothelial hyperplasia, all of which contribute to reduced oxygen tension (Badrah et al., 2022; Li et al., 2022; Li et al., 2023; Rask-Madsen & King, 2013). Although vascular involvement is widely accepted as a primary cause of neuropathy, there is evidence suggesting that these complications may be secondary to a more pronounced neuroglial disorder occurring in anatomical

sites with poor vascularisation (Forbes & Cooper, 2013; Pittenger et al., 2004; Shun et al., 2004).

Structural impairment. Although neuropathy symptoms primarily develop in peripheral structures, evidence indicates that central impairments are already present even with non-neuropathic diabetes and the early stages of diabetic neuropathy. These impairments often coincide with dysfunctions in motor and somatosensory systems, contributing to neuromuscular decline (Allen et al., 2016; Feldman et al., 2019).

Structurally, significant decreases in motor cortex volume have been observed in both T1D (Hughes et al., 2013) and T2D (Peng et al., 2015; Zhang et al., 2022), even in the absence of a neuropathy diagnosis. However, neuropathic patients experience a more severe reduction in motor cortex volume, accompanied by spinal cord atrophy (Selvarajah et al., 2011, 2023; Zang et al., 2023). Structural alterations have also been noted in somatosensory regions, including reduced connectivity between the thalamus and cortex, as well as lower thalamic

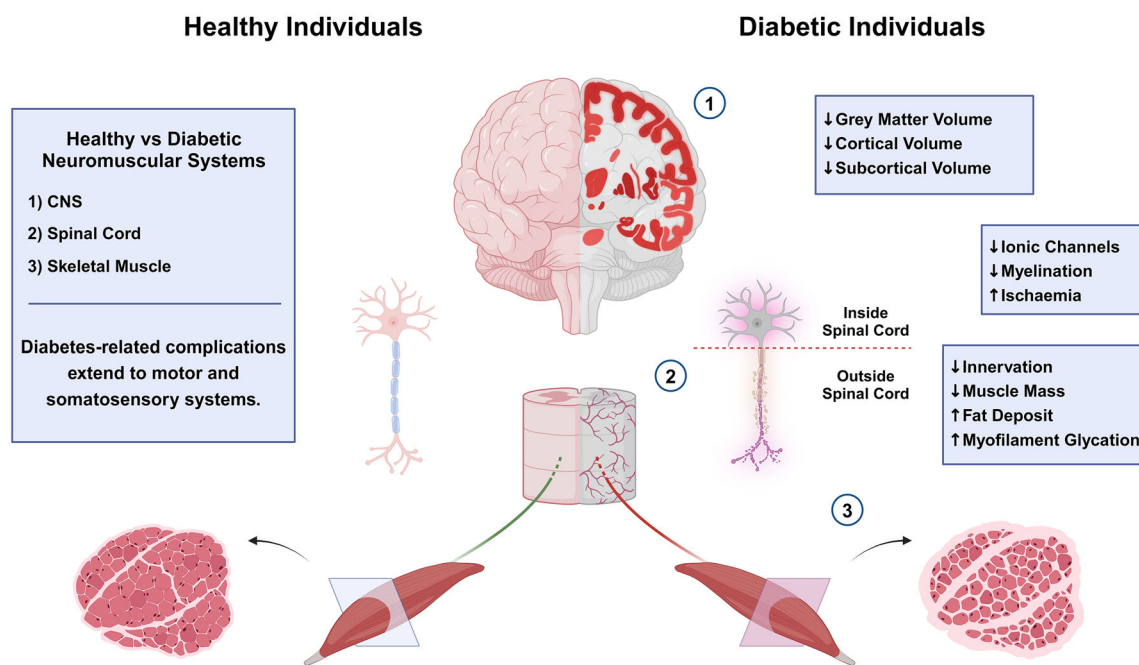


Figure 1. Neuromuscular deterioration observed with diabetes-related complications

(1) In healthy individuals (left), the CNS exhibits adequate grey matter in cortical and subcortical regions. By contrast, diabetic individuals (right) present a significantly lower grey matter volume of cortical and subcortical structures. These alterations suggest neuron and synapse loss alongside disruption of neural networks, potentially impairing cognitive, motor and sensory functions. (2) In healthy subjects, spinal neurons are well-myelinated with adequate ion channels for efficient nerve signal transmission. Myelin facilitates rapid nerve conduction, and good blood perfusion prevents ischaemic phenomena. Diabetic individuals present fewer ion channels and reduced myelination, impairing nerve signal transmission and resulting in slower conduction velocity. Increased ischaemic events indicate reduced blood flow to nerve tissue, exacerbating neuronal damage. (3) Diabetes-related complications impair muscle innervation at the muscular level, reducing contractile capacity and co-ordination. Decreased muscle mass and increased intramuscular fat reflect metabolic imbalance and deteriorated muscle health. Created with BioRender.

grey matter volume in the presence of painful diabetic neuropathy compared to painless neuropathy (Feldman et al., 2019; Selvarajah et al., 2014). Moreover, decreased somatosensory cortex thickness has been reported in both painful and painless neuropathic patients compared to healthy individuals (Selvarajah et al., 2023), with significant axonal degeneration of central somatosensory tracts observed in T2D patients (Fang et al., 2021).

It has also been suggested that vascular damage within the spinal cord, resulting from hyperglycaemia-induced reductions in the vascular endothelial growth factor (VEGF)-A/VEGF receptor 2 signalling cascade, may contribute to neuropathic pain by compromising vascular support to the somatosensory system (Ved et al., 2018). Nonetheless, the precise roles of diabetes vs. ischaemia in these degenerative processes remain unclear, particularly regarding the effects on cerebral neurons and glial cells, including perivascular and mitochondrial swelling (Muramatsu, 2020).

In long-term diabetic patients, spinal motoneurons present severe impairments, including swollen or displaced nuclei and cytoplasmic fat deposits (Reske-Nielsen et al., 1966). These factors significantly contribute to the progression of neuropathy (Andersen, 1999; Said, 2007; Selvarajah et al., 2019). This pathological process, affecting both T1D and T2D individuals, is characterised by motor axon loss, demyelination and neuronal apoptosis (Zhu et al., 2024), resulting in neurogenic muscle atrophy and subsequent declines in motor function (Allen et al., 2013; Allen, Kimpinski, et al., 2014; Picconi et al., 2018; Wong et al., 2017). In addition, spinal neurons exhibit hyperexcitability to peripheral stimuli, with morphological changes such as increased dendritic spine length, head diameter, density and redistribution observed in the presence of diabetic neuropathy, as demonstrated in animal models (Tan et al., 2012). Furthermore, the loss of the motor end plate associated with denervation significantly reduces the number of functional motor units in these individuals (Francis et al., 2011; Muramatsu, 2020). This condition deteriorates in older adults, implicating a severe decline in neuromuscular function as a result of the combined effects of neuropathy and ageing in long-term diabetic individuals (Aagaard et al., 2010; Hepple & Rice, 2016).

Functional impairment. Structural damages typically implicate functional alterations, including deafferentation, impaired somatosensory pathway and reduced thalamic feedback, all contributing to altered pain processing (Feldman et al., 2019). Nociceptive information processing within the spinal cord is regulated by a descending pain modulatory system (Bannister, 2019), where brainstem projections either inhibit or facilitate nociceptive transmission at the spinal level

(Bannister & Dickenson, 2017; West et al., 2015). Dysfunctions in this pathway have been linked to the chronic pain frequently experienced by individuals with neuropathy (Feldman et al., 2019; Ossipov et al., 2014; Tao et al., 2019). The involvement of glial cells and astrocytes in diabetic neuropathy remains poorly understood. However, it has been hypothesised that these cells contribute to altered neural signalling and neuromodulation of somatosensory pathways through the production and release of various chemical factors (Salter & Beggs, 2014).

Altered functionality of somatosensory neurons results in broader input from spontaneously active nociceptors, increased synaptic transmission within the spinal cord and amplified processing of nociceptive signals, complications collectively known as sensitisation (Woolf, 2011; Zang et al., 2023). Impaired sensory feedback probably arises from hyperexcitability or the generation of afferent signals in the absence of external stimuli, leading to dysfunctional stimulus-response coupling (Ørstavik et al., 2006). Hyperexcitability appears to be a primary pathophysiological driver of neuropathic pain (Feldman et al., 2019). This dysfunction is associated with the altered activity of specific sodium and calcium voltage-gated channels involved in nociceptor electrical signalling, as well as the amplification of sensory feedback (Blair & Bean, 2002; Dubin & Patapoutian, 2010; Jagodic et al., 2007; Messinger et al., 2009; Orestes et al., 2013).

Neuropathic complications also include impaired nerve regeneration, primarily as a result of delayed and dysfunctional Wallerian degeneration. Such delays result in reduced reinnervation of target tissues and dysfunctions in neurons and Schwann cells, adversely affecting nerve impulse transmission (Sango et al., 2017; Terada et al., 1998; Yang et al., 2023). A clinical marker of peripheral diabetic neuropathy is reduced nerve conduction velocity (NCV), which is employed to assess the severity of the condition (Lawrence & Locke, 1961; Li et al., 2023; Yorek et al., 2014). Notably, NCV declines progressively with the duration of diabetes and chronic hyperglycaemia in the presence of neuropathy (Feldman et al., 2019; Hamid et al., 2021). The physiological mechanisms underlying reduced NCV include microvascular damage, axonal swelling, dysfunction of the Na^+/K^+ ATPase pump, and altered membrane permeability to sodium and calcium ions (Coppey et al., 2000; Feldman et al., 2019; Forbes & Cooper, 2013; Sima & Brismar, 1985).

Diabetic-related complications of the skeletal muscle

Persistent diabetes leads to impaired skeletal muscle structure and function in both T1D and T2D, with the associated complications progressively reducing

muscle mass, quality and functional capacity. These impairments stem from both neurogenic (neuropathy) and non-neurogenic (myopathy) factors, further damaging motor performance deficits in affected individuals.

Neurogenic disorder in skeletal muscle. The primary cause of reduced muscle mass with diabetes is the accelerated loss of motor axons that mirrors patterns observed with ageing (Allen et al., 2016). This phenomenon arises from cycles of denervation and reinnervation (i.e. sprouting), where certain muscle fibres remain unsuccessfully reinnervated for unknown reasons, contributing to sarcopenia and muscle fibre atrophy (Hepple & Rice, 2016). A more rapid loss of muscle mass probably occurs in both T1D and T2D compared to healthy older adults (Le Corre et al., 2023), who already experience a progressive clustering of fibres within a given motor unit. Therefore, the loss of muscle mass associated with diabetic neuropathy is linked to a dysfunctional sprouting process characterised by accelerated denervation and decreased reinnervation of orphaned muscle fibres (Andersen et al., 1998; Muramatsu, 2020). A lower number of muscle fibres implies a consequent decrease in myosin content, resulting in a significant decline in strength per muscle area, comprising a deconditioning process comparable to that observed in older age and periods of immobilisation (D'Antona et al., 2003).

Non-neurogenic disorder in skeletal muscle. A critical aspect of motor dysfunction is attributed to diabetic myopathy, resulting from multiple physiological determinants, including metabolic and vascular factors (D'Souza et al., 2013; Hernández & Camilo Vanegas, 2015). The progression of diabetic myopathy involves several inter-related mechanisms, including hyperglycaemia-induced oxidative stress, chronic inflammation, excessive fat deposits, decreased mitochondrial density and disrupted extracellular matrix remodelling (Andersen et al., 1998; Hernández & Camilo Vanegas, 2015; Lesniewski et al., 2003; Mesinovic et al., 2019; Perry et al., 2016; Sacchetti et al., 2013).

Chronic hyperglycaemia is associated with the production of reactive oxygen species, leading to mitochondrial dysfunction and promoting a pro-oxidant state that impairs protein synthesis and satellite cell function (D'Souza et al., 2013). This condition can also accelerate muscle protein degradation through the ubiquitin–proteasome pathway, contributing to sarcopenia (Kalyani et al., 2015; Le Corre et al., 2023). Furthermore, persistent hyperglycaemia has been associated with myofibrillar glycation, which implicates a reduced speed of the power stroke. This condition

implicates a structural modification in the catalytic domain of the myosin head observed following *in vitro* exposure to glucose (Ramamurthy et al., 2001, 2003). This complication, combined with impaired microcirculation, compromises the delivery and distribution of metabolic substrates, further worsening the decline in muscle quality (Orlando et al., 2022; Suzuki et al., 2022).

Long-term inflammation is driven by elevated pro-inflammatory cytokines, such as interleukin (IL)-6 and tumour necrosis factor (TNF)- α , which further disrupt muscle function by promoting insulin resistance and reducing regenerative capacity (Hernández & Camilo Vanegas, 2015; Monaco et al., 2019). Furthermore, the accumulation of advanced glycation end-products (AGEs) intensifies inflammation, oxidative stress and muscle degradation, impairing muscle progenitor cell functionality (Hernández & Camilo Vanegas, 2015; Monaco et al., 2019). Increased levels of plasminogen activator inhibitor-1 and collagen are typically observed with diabetes-related complications of the skeletal muscle, implicating disrupted extracellular matrix remodelling and impaired muscle regeneration. In addition, prolonged oxidative stress drives the conversion of muscle stem cells into adipocytes, leading to muscle wasting and metabolic inflexibility (D'Souza et al., 2013; Hernández & Camilo Vanegas, 2015; Monaco et al., 2019).

Although this condition affects both diabetes types, T1D and T2D present a differential diabetic myopathy progression. In T1D, the autoimmune-driven destruction of pancreatic β -cells and subsequent insulin deficiency primarily contribute to the associated complications, which may appear as accelerated muscle ageing with structural changes analogous to those observed in older adults, including mitochondrial dysfunction and muscle atrophy. T2D, by contrast, is often associated with insulin resistance, obesity and increased fat deposition in muscle, leading to metabolic inflexibility and accelerated sarcopenia, especially in older adults (Hernández & Camilo Vanegas, 2015; Monaco et al., 2019). Additionally, T1D-related myopathy often manifests with impaired regenerative responses and reduced muscle satellite cell function, whereas T2D muscles frequently exhibit increased glycolytic fibres and lipid infiltration, contributing to functional decline (D'Souza et al., 2013).

Neuromuscular junction impairments

Motor function decline in diabetic neuropathy is also attributable to dysfunctional neuromuscular junctions (NMJs) (Fig. 2). Age-related NMJ remodelling reduces reinnervation capacity (Pratt et al., 2021) and similar alterations are observed with diabetes, including impaired functionality and a lower number of vesicles

and pre-synaptic mitochondria, as shown in murine models (Estrada-Bonilla et al., 2020; Fahim et al., 1998, 2000; Kimura et al., 1993). As a result, diabetes-related deterioration of NMJ includes diminished calcium sequestration by mitochondria and lower ATP production rate, contributing to impaired excitation–contraction coupling, mirroring NMJ impairments associated with ageing (Anagnostou & Hepple, 2020; Miao et al., 2024; O'Connor et al., 2023). Additionally, lower levels of acetylcholinesterase, which is the enzyme responsible for breaking down ACh outside axon terminals, have been observed in diabetic individuals and correlated with muscle weakness (Garcia et al., 2012).

Post-synaptic alterations include disruptions in nicotinic ACh receptors (nAChRs), resulting in defective ACh–receptor pairing. This disruption weakens the stability and efficiency of neuromuscular transmission, ultimately contributing to reduced motor function (Marques & Neto, 2002). A critical marker of NMJ health is the C-terminal agrin fragment (CAF), a molecule that interacts with the low-density lipoprotein receptor-related protein 4 receptor. Together with the muscle-specific kinase, CAF initiates signalling essential for clustering nAChRs at the post-synaptic membrane, ensuring optimal receptor density and alignment for effective synaptic transmission (Monti et al., 2023). Elevated levels of CAF

in serum reflect NMJ degeneration, indicative of breakdown and reduced function of these synaptic structures. Studies have found these elevated CAF levels in both older adults (Pratt et al., 2022) and individuals with diabetes (Qaisar et al., 2024; Racha et al., 2022), suggesting that NMJ deterioration follows a similar pattern in both conditions. This observation reinforces the concept of diabetes-related neuromuscular complications as a model of accelerated ageing because NMJ function diminishes more rapidly and parallels age-related degeneration.

Neural control with diabetic neuropathy. Motor control is accomplished when descending neural signals elicit co-ordinated muscle responses to generate planned movements. Afferent and efferent signals integrate at both spinal and supraspinal levels, influencing the output of spinal motoneurons to muscle fibres (Avaltroni et al., 2024; Gandevia, 2001; McLean et al., 2007; Osseward & Pfaff, 2019). Upon reaching the motor end plate, these signals trigger calcium release from the sarco/endoplasmic reticulum, prompting muscle fibres to generate tension (Calderón et al., 2014). A single motoneuron and the muscle fibres it innervates represent the motor unit, which is recognised as the final common pathway for motor command (Duchateau & Enoka, 2011; Liddell & Sherrington, 1925;

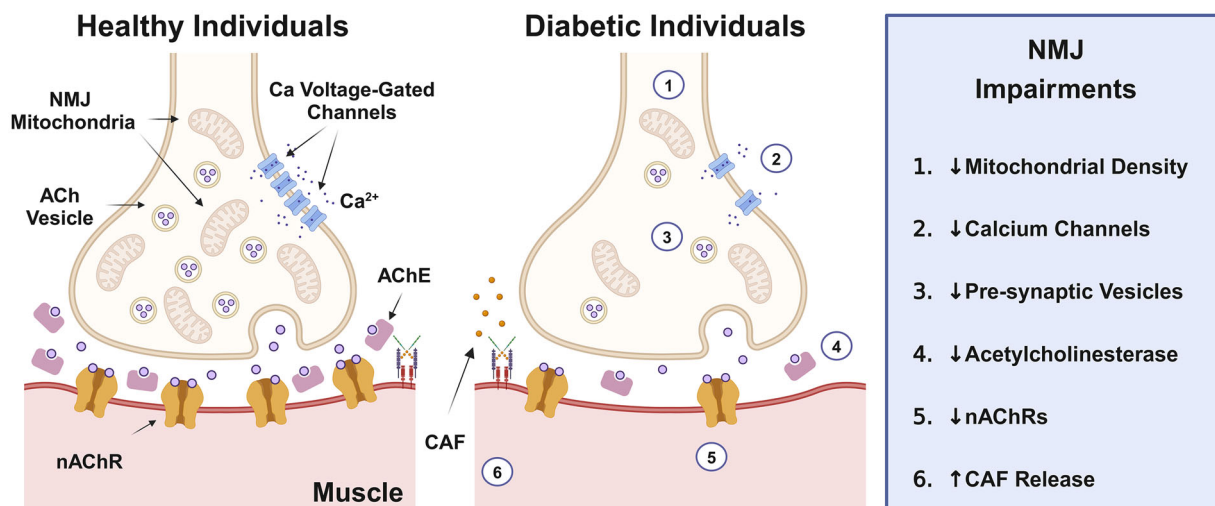


Figure 2. Neuromuscular junction deterioration in diabetic individuals

In healthy individuals (left), neuromuscular junctions show a well-organised structure with a homogeneous distribution and a high density of post-synaptic nicotinic ACh receptors. The abundant presence of calcium channels and pre-synaptic vesicles ensures efficient synaptic transmission together with a proper mitochondrial density, supporting energy demands and calcium buffering in the axonal terminal. Acetylcholinesterase activity allows rapid degradation of ACh, maintaining synaptic balance and optimal muscle function. In diabetic individuals (right), neuromuscular junction (NMJs) are significantly impaired, presenting lower mitochondrial density (1), limiting energy production capacity and calcium sequestration. Decreased calcium channels (2) and pre-synaptic vesicles (3) further impair neuromuscular transmission. Lower acetylcholinesterase content (4) reduces the ACh degradation rate, impairing proper reuptake. Furthermore, a lower density of post-synaptic nicotinic ACh receptors (5) implicates decreased vesicle-releasing-binding coupling. The increased release of C-terminal agrin fragment (CAF) is used as a serum biomarker of NMJ deterioration (6). AChE, acetylcholinesterase; Ca^{2+} , calcium (ion); nAChR, nicotinic ACh receptor.

Sherrington, 1906). Therefore, motor output is shaped by multiple inputs converging on spinal motoneurons, including neural feedback from receptors and signals from spinal and supraspinal areas (Farina & Gandevia, 2024; Gandevia, 2001; Gandevia et al., 1990; Heroux et al., 2022; Proske & Gandevia, 2012). As a result, neuromuscular function relies on co-ordinated motor unit activation and firing patterns to ensure precise control of muscle contraction (Enoka & Fuglevand, 2001; Heckman & Enoka, 2012; Henneman, 1957). Extensive research has demonstrated that this finely tuned system is severely compromised by diabetic neuropathy, resulting in numerous pathophysiological consequences that affect both the motor and somatosensory nervous systems (Andersen et al., 1996, 2004, 2012; Hilton et al., 2008; Ho et al., 2013; Malone, 2016; Muramatsu, 2020).

Diabetes-related complications on motor unit function.

Recording motor units offers valuable insights into motoneuron properties (Farina & Gandevia, 2024). This is accomplished by analysing individual motor unit activity, reflecting the one-to-one association between action potentials generated by an alpha motoneuron and those received by its innervated muscle fibres (Martinez-Valdes et al., 2023). Such recordings facilitate the understanding of the peripheral motor control deterioration observed in diabetic individuals. Impaired motor unit characteristics include suppressed firing rates, reduced post-activation potentiation (Allen et al., 2016) and inefficient recruitment during sustained contractions, where the same limited pool of motor units is repeatedly activated (Watanabe et al., 2012).

Evidence obtained using high-spatial resolution electromyography confirmed lower discharge rates, marked by higher firing fluctuations and impaired force steadiness in neuropathic individuals compared to healthy controls (Favretto et al., 2023; Senefeld et al., 2020). More recently, reduced firing rates during recruitment and sustained contraction phases have also been observed in young individuals with T1D compared to healthy individuals despite comparable maximal voluntary force levels (Valli et al., 2025). However, a meta-analysis discussing EMG studies evaluating lower limb function with diabetic neuropathy revealed significant result heterogeneity, highlighting the need for further research to fully elucidate the physiological mechanisms underlying motor impairments across different stages of neuropathy (Haque et al., 2020).

The effect of diabetes-related complications on performance decline. The neurogenic deterioration of skeletal muscle quality is widely recognised as the primary factor contributing to impaired neuromuscular function in diabetic neuropathy (Parasoglou et al., 2017). The

degenerative processes associated with diabetes-related complications are significantly intensified in the presence of neuropathy. The resulting pathological alterations lead to a dysfunctional integration of somatosensory inputs and motor responses, culminating in impaired motor unit functioning (Allen, Kimpinski, et al., 2014; Favretto et al., 2023; Le Corre et al., 2023; Watanabe et al., 2012). This disruption of the motor-sensory system initiates a cascade of complications that progressively deteriorate neurophysiological function, contributing to neuromuscular performance decline. Such a decline is particularly critical for maintaining motor function and independence in individuals with long-term diabetes, who also experience age-related physiological deterioration (Forbes & Cooper, 2013).

Muscle strength, power and rate of force development.

Impaired neuromuscular function in diabetes is associated with a suppressed force generation capacity, ~20% lower in uncomplicated diabetic individuals compared to healthy counterparts (Andersen et al., 1996; Orlando et al., 2016). This impairment is attributed to diminished muscle quality in both T1D and T2D, resulting in a progressive and accelerated decline in muscle strength, reaching an up to ~40% decrease in neuropathic individuals compared to healthy counterparts (Andreassen et al., 2006; Le Corre et al., 2023). Non-neuropathic diabetic individuals present ~40% lower muscle power and rate of force development (RFD) than healthy controls (Favretto et al., 2019; Sacchetti et al., 2013; Senefeld et al., 2020). These reductions are even more severe in neuropathic patients, with an ~70% lower power output and an RFD reduced by ~50% (Favretto et al., 2019; Hilton et al., 2008; Le Corre et al., 2023).

Muscle endurance. Diabetes and associated complications negatively affect muscle endurance, inducing significant decreases in the time to task failure in both types of diabetes (T1D: ~30% in mixed samples; T2D: ~30% in males and ~20% in females) and larger force decrease (i.e. fatigue index) (both T1D and T2D: ~20%) compared to healthy counterparts (Bazzucchi et al., 2015; Orlando et al., 2017, 2017a, 2017b). Individuals with neuropathic complications exhibit lower muscle endurance and higher force decrease, ~20% and 35% lower than in non-neuropathic T1D (Orlando et al., 2017b) and T2D (Orlando et al., 2022). Furthermore, neuropathic individuals exhibit ~50% lower muscle endurance than healthy controls (Orlando et al., 2017b). The evidence presented above indicates that T1D and T2D result in a comparable decline in performance, whereas a more severe loss of function is observed with neuropathy (Fig. 3). This decline is further exacerbated by ageing in diabetic individuals, with performance reducing

up to 50% more than in healthy ageing, according to longitudinal investigations (Seok et al., 2007).

Physiological determinants of neuromuscular performance decline. Reduced performance is hypothesised to be influenced by preferential denervation of type II muscle fibres (Allen, Major, et al., 2014; Le Corre et al., 2023), although neural impairments also play a significant role. Nevertheless, RFD relies on both neural and contractile properties (Del Vecchio et al., 2024; Maffiuletti et al., 2016), with the speed of motoneuron recruitment and maximal discharge rate being essential for rapid force generation (Del Vecchio et al., 2019), both of which are compromised with neuropathic and non-neuropathic diabetes. A possible explanation for

the pronounced decline in RFD and power output is the lower muscle fibre conduction velocity observed in diabetic individuals (Hilton et al., 2008; Sacchetti et al., 2013). Furthermore, the greater reduction in muscle power compared to RFD is attributable to impaired cross-bridge kinetics and reduced muscle shortening speed in diabetes. Power is assessed during dynamic contractions, whereas RFD is typically measured during isometric contractions (Le Corre et al., 2023; Sacchetti et al., 2013). The decline in muscle power precedes that of force in the presence of diabetes, a condition that worsens with ageing, long-term diabetes and neuropathy (Hepple & Rice, 2016; Le Corre et al., 2023; Lesniewski et al., 2003; Sacchetti et al., 2013). These findings underscore the need for further research into the effects of physical activity on





| | Diabetes (Non-Neuropathic) | | Diabetes (Neuropathic) | |
|--|----------------------------|--|------------------------|---|
| | Decline | Physiological Mechanisms | Decline | Physiological Mechanisms |
|  Maximal Strength | -20% | ↓ Volitional Activation ↓ Cross-Bridge Force ↓ Muscle Mass | -40% | ↓ Glial Cell Functioning ↑ Spinal Cord Damage ↓ Type II Muscle Fibres |
|  Maximal Power | -40% | ↓ Volitional activation ↓ NMJ transmission ↓ Cross-Bridge Force ↓ Muscle Mass | -70% | ↓ MNCV ↑ Spinal Cord Damage ↓ Recruitment Speed |
|  RFD | -40% | ↓ Volitional activation ↓ NMJ transmission ↓ EC coupling ↓ Muscle Mass | -50% | ↓ MNCV ↑ Spinal Cord Damage ↓ Recruitment Speed |
|  Muscle Endurance | -30% | ↓ Mitochondrial Density ↓ Contractile Properties ↑ Myofilament Glycation | -50% | ↑ Hypoxia ↑ Pain perception ↑ Chronic inflammation |

Figure 3. Performance decline with diabetes-related complications
 Four aspects of muscular performance, (1) maximal strength, (2) maximal power, (3) rate of force development (RFD) and (4) muscle endurance, are primarily impaired in diabetic individuals. (1) Maximal strength is reduced by 20% in diabetes as a result of decreased grey matter, lower muscle activation and muscle mass. In diabetic neuropathy, strength reduction reaches 40%, attributed to glial cell dysfunction, spinal cord damage, and type II fibre loss. (2) Maximal power decreases by 40% in diabetes, linked to lower muscle activation, neuromuscular junction (NMJ) transmission and excitation–contraction (EC) coupling; this reduction reaches 70% with neuropathy as a result of slower motor unit recruitment and reduced nerve conduction. (3) RFD is reduced by 40% in diabetes and by 50% in neuropathy, with the latter linked to more severe spinal damage and slower motor recruitment. (4) Muscle endurance is lowered by 30% in diabetes as a result of decreased mitochondrial density, contractile properties and myofilament glycation, whereas neuropathy results in a 50% decrease, exacerbated by hypoxia, pain and chronic inflammation. The data compare affected adults to healthy controls. MNCV, motor nerve conduction velocity. Data from Allen et al. (2015, 2016); Allen, Major et al. (2014); Bazzucchi et al. (2015); Hilton et al. (2008); and Orlando et al. (2016, 2017b, 2017a).

Table 1. Mechanisms of central and peripheral fatigue with diabetes-related complications

| | Determinant | Impact on fatigue |
|-----------------------|----------------------------|-------------------------------|
| Central mechanisms | ↓ Grey matter volume | ↓ Motor control |
| | ↓ Neural connectivity | ↓ Neural input integration |
| | ↑ Spinal cord vasculopathy | ↓ Motor co-ordination |
| Peripheral mechanisms | ↑ Spinal nerve atrophy | ↓↑ Fatigue perception |
| | ↓ NMJ function | ↓↑ Spinal signalling |
| | ↓↑ Pain perception | ↓ Motoneuron activation |
| | ↓ Mitochondrial density | ↓↑ Neural transmission |
| | ↓ Muscle vascularisation | ↓↑ Integration-response tasks |
| | ↑ Myofilament glycation | ↓↑ Neuromuscular transmission |
| | ↓ Calcium availability | ↑ Discomfort |
| | | ↑ Perceived fatigue |
| | | ↓ ATP availability |
| | | ↑ Early fatigue onset |
| | ↑ Oxygen flow | |
| | ↓ Metabolite disposal | |
| | ↓ Force output | |
| | ↑ Effort | |
| | ↓↑ E-C coupling | |

↓, reduced; ↑, increased; ↓↑, altered; E-C, excitation-contraction; NMJ, neuromuscular junction.

the progression of diabetes-related complications and the implication for mitigating symptoms and performance decline.

Mechanisms of fatigue in diabetic-related complications

It is essential to differentiate exercise-induced fatigability from the trait fatigue commonly associated with diabetes. Exercise-induced fatigability results from temporary, task-driven muscle exhaustion. Trait fatigue is a persistent fatiguing state influenced by glycaemic dysregulation, depression and lifestyle (Behrens et al., 2023). Although exercise-induced fatigability occurs during physical exertion and primarily depends upon physiological training conditioning, trait fatigue persists regardless of physical activity and can significantly impact daily functioning and quality of life (Behrens et al., 2023; Fritschi & Quinn, 2010; Kalra & Sahay, 2018). In particular, task-induced fatigue encompasses central and peripheral mechanisms that influence both motor performance and perceived dimensions.

Motor performance fatigue is defined as a measurable decline in motor output, such as reduced maximal voluntary contraction or decreased endurance, attributed to impairments within the neuromuscular system. This type of fatigue is objective, resulting directly from physiological limitations in neural and muscular functions. By contrast, perceived motor fatigue refers to the subjective sensation of tiredness or a need to reduce effort, influenced by psychophysiological factors such as mood, motivation

and discomfort signals from sensory feedback (Behrens et al., 2023). In the presence of diabetes, in particular, diabetic neuropathy, these conditions are typically interdependent as a result of psychophysiological decline, nerve dysfunctions and cardiometabolic impairments (Fritschi & Quinn, 2010; Kalra & Sahay, 2018).

Central and peripheral mechanisms of fatigue do not operate in isolation but instead interact in a way that intensifies the overall experience of fatigue in diabetic individuals (Table 1). Central deficits impair motor control and signal co-ordination, placing greater strain on the peripheral musculature to maintain performance. Simultaneously, peripheral muscle dysfunctions, including reduced mitochondrial function, impaired blood flow and structural abnormalities, feedback to the CNS through altered sensory inputs. This feedback can exacerbate perceived fatigue, leading to an early sensation of exhaustion, further intensified by altered pain perception (Allen et al., 2015; Feldman et al., 2019; Fritschi & Quinn, 2010).

Central mechanisms of fatigue. Central fatigue in diabetes and diabetic neuropathy primarily results from neurological impairment that affects neural transmission, co-ordination and motor control (Allen et al., 2015; Fritschi & Quinn, 2010). Reduced grey matter volume in the cortex, commonly observed in individuals with diabetes, diminishes voluntary activation and the capacity of the CNS to perform sustained, high-intensity tasks (Fritschi & Quinn, 2010; Muramatsu, 2020). This impairment limits cognitive-motor integration,

which is critical for maintaining optimal motor output. Additionally, decreased connectivity between cortical and subcortical areas weakens the communication between regions responsible for motor planning, control and feedback (Fritschi & Quinn, 2010; Kalra & Sahay, 2018). Furthermore, the efficiency in motor unit recruitment from the CNS is reduced as a result of a detrimental synchronisation in these brain networks, leading to a faster onset of fatigue and a reduced ability to sustain motor performance over time (Fritschi & Quinn, 2010; Kalra & Sahay, 2018; Watanabe et al., 2012).

Further complicating these central mechanisms, spinal cord vasculopathy and atrophy compromise blood flow and nutrient delivery to spinal neurons, which are essential for maintaining motor unit recruitment during physical activity (Allen et al., 2015; Ved et al., 2018). With diabetic neuropathy, this spinal cord impairment affects both the neural transmission along the spinal pathways and the feedback mechanisms that normally regulate motor unit activation (Muramatsu, 2020). As a result, the CNS is forced to increase cortical activation for motor output, which is inherently unsustainable over prolonged periods. Diabetic neuropathy also introduces specific disruptions in motor unit recruitment patterns, with irregular motor unit firing, a condition requiring increased energy expenditure to achieve consistent force output (Favretto et al., 2019, 2023; Watanabe et al., 2012). This dysfunction further amplifies central fatigue as the CNS struggles to co-ordinate and sustain motor output in a condition of irregular and inefficient neural signalling.

Peripheral mechanisms of fatigue. Structural and functional impairments contribute to peripheral fatigue as a result of diabetes-related complications, particularly at the muscle fibres and NMJ level. Sensory and motor nerve atrophy, along with the degeneration of motor end plates, significantly affects neuromuscular transmission (Allen et al., 2015; Le Corre et al., 2023). Loss of motor end plate integrity means that, even with adequate neural drive, the muscular response is compromised, accelerating fatigue onset during physical exertion. Compounded by impaired NMJ function, the transmission of action potentials to muscle fibres is weakened, further impacting excitation–contraction coupling (Anagnostou & Hepple, 2020; Miao et al., 2024; Orlando et al., 2022). These complications contribute to performance fatigue as muscle fibres become less responsive to neural signals, necessitating greater effort to sustain contraction and maintain performance.

Peripheral fatigue is also marked by myofibrillar glycation, a process dependent on chronic hyperglycaemia that structurally alters myosin filaments within muscle fibres (Ramamurthy et al., 2001, 2003). This glycation reduces the efficiency of cross-bridge cycling, leading

to a weakened and slower rate of force production and compelling the body to expend additional energy for the same mechanical output level (Orlando et al., 2022). Dysfunctional excitation–contraction coupling, partly as a result of calcium release abnormalities in the sarcoplasmic reticulum, compounds these peripheral limitations by further reducing contractile efficiency, thereby accelerating fatigue onset (Orlando et al., 2022).

Compromised supply and health of muscle fibres observed with diabetic neuropathy further exacerbate peripheral fatigue, with vascular damage impacting perfusion, oxygen tension (Li et al., 2023) and reduced mitochondrial density, limiting ATP production necessary for muscle contractions (Andersen, 1999; Li et al., 2023). Energy sources become insufficient to meet performance demands, leading to early muscle fatigue as energy reserves are depleted, particularly during prolonged tasks.

Together, these peripheral impairments form a substantial barrier to sustained muscle activity because both structural and metabolic complications undermine the muscular ability to maintain prolonged contractions.

Perceived fatigue in diabetic individuals. Perceived fatigue reflects the subjective sense of exhaustion or weariness, which can arise independently of physical exertion (Behrens et al., 2023). In the presence of diabetes, perceived fatigue significantly limits physical performance because it is heavily influenced by the underlying pathophysiology of diabetes-related complications. The combination of metabolic, sensory and emotional factors creates both physical and cognitive experiences of fatigue, disrupting motor performance and exercise tolerance (Kalra & Sahay, 2018). Unlike performance fatigue, which is directly measurable through objective metrics, perceived fatigue in diabetic individuals closely aligns with the subjective feeling of energy depletion, often resulting in a mismatch between perceived effort and actual performance levels (Barakou et al., 2023).

Neuropathic complications further exacerbate this issue by disrupting afferent signals that communicate muscle status to the CNS, leading to an amplified sense of effort, particularly in those with diabetic neuropathy (Barakou et al., 2023; Fritschi & Quinn, 2010). These sensory disruptions increase the perceived effort for simple motor tasks, such that even daily activities may feel as demanding as moderate to intense exercise. This altered sensory feedback affects proprioceptive inputs, increasing the subjective experience of fatigue and lowering the threshold for exhaustion during physical exertion.

Psychological factors also contribute to higher perceived fatigue in diabetic patients, influencing cognitive and physical domains. Diabetes-related psychological distress, encompassing diabetes distress,

anxiety and depression, is prevalent in this population, further increasing perceived fatigue (Kalra & Sahay, 2018). The chronic mental load required to manage blood glucose, dietary restrictions and medication schedules results in persistent cognitive fatigue (Barakou et al., 2023; Fritschi & Quinn, 2010), directly impacting motor performance by reducing the individual's mental resilience and the ability to sustain effort during exercise.

Older females with diabetes generally report higher perceived exertion than males in comparable conditions (Huebschmann et al., 2015). Furthermore, in controlled exercise settings, such as endurance step tests at 50–90% of the maximal workload, no significant differences have been reported in male diabetic individuals (50–60 years of age, non-neuropathic) in terms of both absolute heart rate and heart rate relative to the maximum. Similarly, no differences emerged in perceived exertion across each test level for these male participants (Schwensfeier et al., 2023). Discrepancies in perceived fatigue across studies probably arise from variations in testing conditions, demographics and individual health factors. Sex-specific physiological and psychological differences and distinct study designs may influence the mentioned outcomes. Moreover, the diverse impact of diabetes-related complications such as neuropathy and individual approaches to diabetes management suggest the need for additional research for a comprehensive characterisation of precise determinants of perceived fatigue in this population.

Glycaemic control and the protective role of physical activity

Managing blood glucose levels prevents and mitigates micro- and macrovascular complications in diabetic individuals (Le Corre et al., 2023). Dysglycaemia is associated with a higher risk of developing endothelial dysfunctions, contributing to a greater incidence of cardiovascular complications (Li et al., 2023). Both long-term and short-term glycaemic control are crucial in supporting cardiovascular health and preventing complications such as diabetic neuropathy (Feldman et al., 2019; Forbes & Cooper, 2013).

Impaired long-term glucose control is responsible for adverse cardiometabolic events, commonly recognised as glucotoxicity (Rossetti et al., 1990), implicating consequences such as reduced insulin sensitivity and secretion, as well as decreased β -cell mass and functionality (Bensellam et al., 2012; Robertson et al., 2003). These mechanisms progressively lead to insulin resistance and impaired glucose uptake in T2D individuals. Moreover, glucotoxicity is associated with increased oxidative stress, cellular damage, endothelial dysfunction and inflammation, heightening the risk of

both microvascular and macrovascular complications in individuals with either T1D or T2D (Le Corre et al., 2023; Li et al., 2023).

It is fundamental to note that other factors may influence the appearance of microvascular complications in diabetic individuals. Short-term glycaemic variability is typically associated with an increased risk of developing neuropathic complications (Zhang et al., 2021). In particular, individuals with diabetic neuropathy have demonstrated higher variability in glycosylated haemoglobin (HbA1c) compared to those without complications (Su et al., 2018). Daily fluctuations in glucose levels contribute to increased oxidative stress and elevated circulating inflammatory factors (e.g. IL-6, TNF- α and IL-18), as well as adhesion molecules (intercellular adhesion molecule-1, vascular cell adhesion molecule-1 and E-selectin), collectively raising the incidence of cardiovascular complications in both types of diabetes (Ceriello et al., 2004, 2010; Ceriello & Ihnat, 2010).

Postprandial periods amplify the consequences of glycaemic variability, with glucose levels often exceeding normal ranges. This underscores the relevance of managing post-meal glucose levels to improve overall cardiometabolic health in diabetic individuals and reduce complications (Ceriello et al., 2004). In these terms, exercise and diet are essential for reducing the post-meal glucose response and glycaemic variability, lowering the risk of developing vascular complications (Bellini et al., 2024; Syeda et al., 2023; Yorek et al., 2014; Zhang et al., 2021). For example, it has been recently highlighted that prolonged moderate-intensity exercise starting soon after eating (i.e. 30–60 min after) can significantly reduce the postprandial glucose peak (Bellini et al., 2021). Therefore, implementing physical activity right after a meal would benefit cardiometabolic health.

Technological advances are also helpful in glycaemic management, particularly through continuous glucose monitoring systems. These systems allow diabetic individuals to more effectively manage glucose fluctuations, increasing time within the target glycaemic range and decreasing daily variability (Yapanis et al., 2022). Additionally, continuous glucose monitoring is helpful in monitoring glucose levels during exercise, allowing real-time adjustments that enhance exercise safety. This capability may improve adherence to exercise programs because fear of hypoglycaemic events during physical activity is a common reason for exercise avoidance in diabetic populations (Cigrovski Berkovic et al., 2021; Zaharieva & Addala, 2022).

The benefits of exercise in diabetic individuals extend beyond managing glucose and lipid metabolism, and diverse physiological adaptations result from long-term exercise programs (Praet et al., 2007). Both resistance and endurance exercises have been associated with beneficial effects for neuropathic symptoms, glycaemic control,

and physical fitness (Li et al., 2024; Pedersen, 2017; Su et al., 2023). These beneficial effects are achieved through specific physiological responses to different physical training modalities summarised in Fig. 4.

Aerobic training. People diagnosed with diabetes mellitus typically present a lower exercise capacity by

~20% (Wei et al., 1999; Zaccardi et al., 2015), manifested with a reduced $\dot{V}_{O_{2max}}$ and time-to-exhaustion compared to healthy, age and physically-activity-matched control groups (Nesti et al., 2020). Decreased exercise capacity includes deficiencies in the cardiovascular and respiratory systems to supply oxygen, as well as the inability of tissues to extract oxygen (Nesti et al., 2020; Reusch et al.,

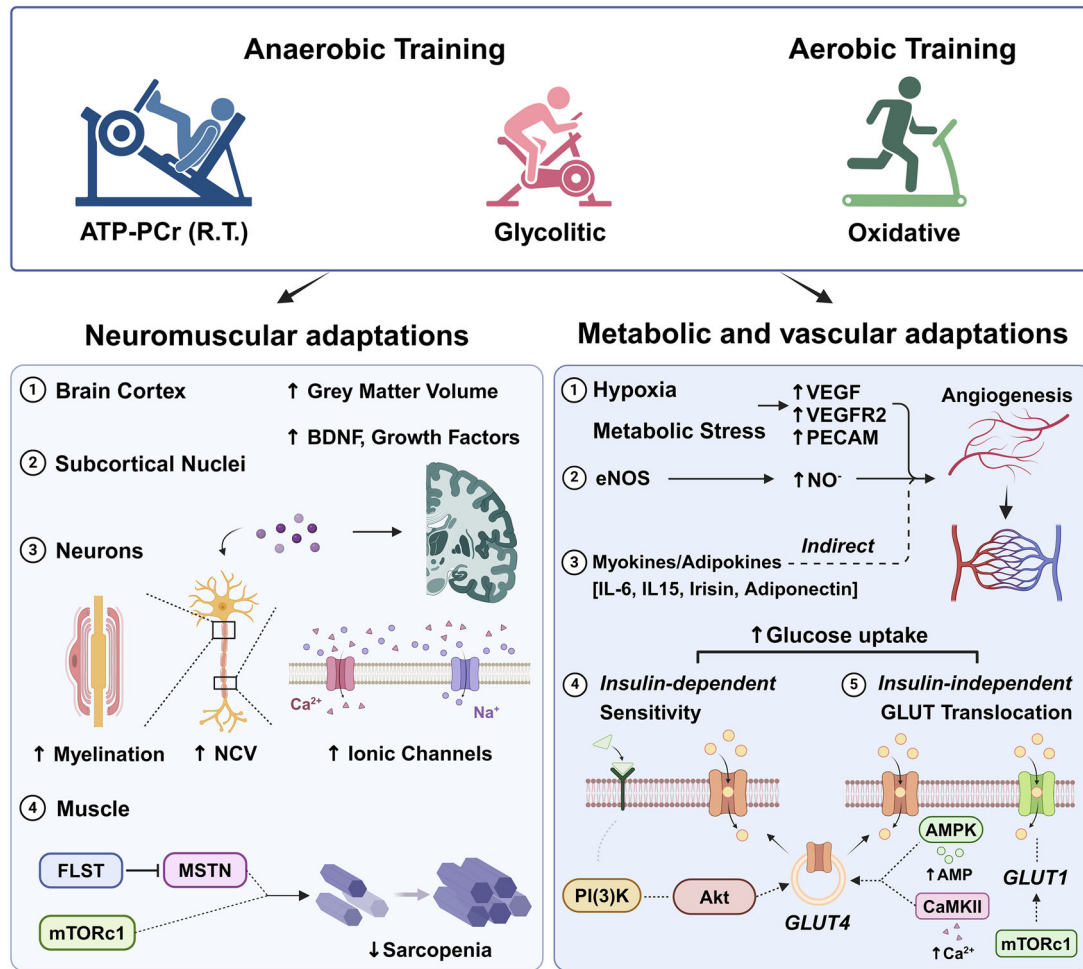


Figure 4. Physiological and molecular changes induced by physical exercise

Both anaerobic (i.e. resistance training/ATP-PCr, glycolitic energy systems) and aerobic (oxidative energy system) training promote neuromuscular (right) and metabolic/vascular adaptations (left). Right: the neuromuscular effects of exercise highlight changes in the central and peripheral nervous system. Exercise promotes the increase in grey matter and brain-derived neurotrophic factor (BDNF) in the cerebral cortex (1) and subcortical nuclei (2), as well as an increase in other beneficial growth factors. The release of these substances at the neuronal level (3) induces an improvement in myelination, an increase in nerve conduction velocity (NCV) and an enhancement of ion channels. As a result of the combined effect of mechanical overload and time under tension, molecular signalling of mechanistic target of rapamycin complex 1 (mTORc1) and follistatin (inhibiting myostatin), exercise reduces sarcopenia and improves muscle function. Left: exercise-induced hypoxia (1) and metabolic stress (2) lead to a significant increase in the expression of angiogenic factors, such as vascular endothelial growth factor (VEGF), VEGFR2 (VEGF receptor) and endothelial plaque cell adhesion molecule (PECAM), as well as endothelial nitric oxide synthase and nitric oxide (2). These changes and the secretion of myokines and adipokines (3) facilitate angiogenesis, improving blood flow and nutrient supply to muscle tissue. In addition, there is an improvement in glucose uptake mediated by both insulin-dependent (4) and insulin-independent glucose transporter type 4 (GLUT-4) receptors (5), which contributes to increased energy availability and optimised metabolic function. AMPK, adenosine monophosphate kinase; eNOS, endothelial nitric oxide synthase; FLST, follistatin; IL, interleukin; MSTN, myostatin; NO, nitric oxide; PCr, phosphocreatine; R.T., resistance training. Created with BioRender.

2013). Endurance training is considered safe and effective in enhancing glycaemic control, exercise capacity, and well-being status in complicated and uncomplicated diabetic individuals (Colberg et al., 2016; Kanaley et al., 2022). Aerobic exercise promotes the decrease in HbA1c and fasting plasma glucose levels (Boulé et al., 2003; Umpierre et al., 2011) in a dose-dependent association with modifications in training volume and/or intensity (Delevatti et al., 2019).

Improvements in aerobic exercise capacity are mainly related to modifications in the oxygen cascade, with an augmented capacity of oxygen transportation and extraction observed in diabetic individuals (Nesti et al., 2020). For example, a study investigating cardiac modifications following a high-intensity interval training program has shown that diabetic individuals had greater cardiac output at rest, which was accompanied by improvement in left ventricular filling and emptying capacity, as well as an increase in stroke volume (Wilson et al., 2019). Nevertheless, it has been shown how both continuous and interval training were effective strategies to improve oxygen extraction in the diabetic population (Van Ryckeghem et al., 2022) and these findings pointed out reduced blood flow to skeletal muscle as a contributing factor to exercise tolerance in T1D and T2D patients (Goulding et al., 2020; Nesti et al., 2020).

Despite vascular damage induced by chronic hyperglycaemia, it has been suggested that aerobic exercise performed at both moderate and high intensity promotes angiogenesis, decreases capillary rarefaction and reduces pericapillary membrane thickness (Basu et al., 2014; Mortensen et al., 2019). Among several underlying mechanisms involving glycaemic and AGEs control, inflammation management, and apoptosis regulation (Callaghan & Feldman, 2013; Dixit et al., 2014; Gholami et al., 2018; Olver et al., 2014), reactive oxygen species, myokines and the upregulation of endothelial nitric oxide synthase promoting nitric oxide (NO⁻) synthesis are the leading candidates inducing vascular adaptations in response to exercise (Groen et al., 2014; Prior et al., 2014; Qi et al., 2022; Song et al., 2024). Nevertheless, these findings remain highly speculative in people affected by diabetes mellitus because these individuals typically present NO resistance (Bahadoran et al., 2023).

However, the development of microvascular complications with diabetes is associated with increased production of reactive oxygen species (Baynes, 1991; Giugliano et al., 1996), disrupting the balance between free-radical generation and antioxidant activity (Giacco & Brownlee, 2010). A long-term endurance training program has been associated with decreased oxidative stress among people with T2D by modifying the systemic oxidative stress profile (Nojima et al., 2008) and selectively upregulating antioxidant enzymes compared to other training modalities (de Oliveira et al., 2012).

Aerobic training effectively reduces fat content in diabetic patients, as demonstrated by reductions in visceral and subcutaneous adipose tissue following an aerobic training program (Bacchi et al., 2012). These adaptations were also observed in another study on diabetic individuals (Seung et al., 2005), outlining modifications in both absolute and relative terms. Altered fat mass in diabetic individuals may result from changes in cytokine levels (i.e. TNF and IL-6), which commonly have inhibitory effects on insulin secretion, leading to insulin resistance (Hajer et al., 2008). Indeed, higher levels of IL-6 are typically observed after acute exercise, which suppresses the production of TNF and enhances insulin secretion with a concomitant effect on glycaemic function through stimulation of glucagon-like peptide-1 from pancreatic β -cells (Karstoft & Pedersen, 2016). Constant and regular exercise practice indirectly affects chronic inflammation through improvements in body composition, physical fitness and metabolic function (Colberg et al., 2016; Pedersen & Saltin, 2015), modulating the activity of pro-inflammatory cytokines (Papagianni et al., 2023). Moderate-intensity aerobic training programs promote reductions in C-reactive protein (Balducci et al., 2010; Choi & Rush, 2012) and resistin levels (Giannopoulou et al., 2005), as well as IL-6 and TNF, as previously outlined. The above findings point out the effects of regular endurance training on the damage exerted by chronic hyperglycaemia in diabetic individuals.

Moreover, aerobic training promotes the activation of PGC-1 α , which is critical in mediating mitochondrial biogenesis and neuronal cell survival (Halling & Pilegaard, 2020; You et al., 2024). These adaptations have been observed in people with both types of diabetes in response to various aerobic training modalities, even though to a lower extent than in healthy individuals (Jingjing et al., 2023; Lin et al., 2024; Minnock et al., 2022), implicating greater oxidative capacity of the skeletal muscle (Lundby & Jacobs, 2016). Mitochondrial function enhancements have been observed in response to moderate-intensity endurance training, evidenced by a higher creatine phosphate recovery rate. This improvement is further supported by elevated insulin-stimulated glucose oxidation and reduced fat oxidation (Meex et al., 2010). Another study examining the effects of low-volume, high-intensity interval training reported increased citrate synthase activity alongside other skeletal muscle mitochondrial capacity markers. However, when comparing moderate- and high-intensity training programmes in diabetic mice, no significant differences in mitochondrial function were found (Chavanelle et al., 2017). These findings, however, await confirmation in human studies.

It has been reported that 6 months of aerobic exercise mitigates the deterioration of brain structures in the T2D

population. In particular, a lesser duration of physical activity (hours/week) is associated with a lower volume of cortical and subcortical structures, which is not dependent on HbA1c (Moreno et al., 2023). Furthermore, diabetic individuals with and without neuropathy undertaking aerobic exercise significantly improved NCV (Balducci et al., 2006; Gholami et al., 2018), leading to enhanced functionality in both sensory and motor nerves because of the positive effect of exercise (Beigi et al., 2023; Streckmann et al., 2022). These findings support the notion of physical training safety and effectiveness in restoring neural functioning in diabetic individuals with neuropathic complications. Moreover, aerobic exercise promotes several *exerkines*, such as irisin, adiponectin and interleukins, which have been recognised as a primer for molecular signalling of injured tissue regeneration (Chen et al., 2022).

Aerobic exercise also induces motor cortical neuroplasticity in diabetic individuals, comprising a critical adaptation in mitigating neural deterioration occurring with neuropathic complications (McDonnell et al., 2013; Yamazaki et al., 2019; Yang et al., 2019). This is probably induced by the exercise role in promoting brain-derived neurotrophic factor (BDNF), which is responsible for neuronal survival, proliferation, maturation, and outgrowth in both the central and peripheral nervous systems, including beneficial effects for spinal cord injuries (Bilchak et al., 2021; Cefis et al., 2023; Chang et al., 2019; Chen et al., 2022; English et al., 2014; McGregor & English, 2019).

To date, no studies assessing NMJ modification following exercise intervention have been performed in diabetic individuals. However, healthy young and older adults exhibited enhanced pre-synaptic nerve terminal branching and the number of vesicles, along with a greater number of post-synaptic receptors, with significant responses observed after endurance exercise (Deschenes, 2019; Yamaguchi et al., 2024).

Resistance training. The increases in muscle strength in response to resistance training are accompanied by a higher muscle fibre conduction velocity in diabetic individuals, indicative of enhanced recruitment of higher-threshold motor units (Bazzucchi et al., 2015). These adaptations are particularly evident in lower limbs compared to upper limbs as a result of differences in fibre morphofunctional features, as well as sarcolemmal excitability (Orlando et al., 2016). In this context, performance enhancements from training can result from spinal and supraspinal adaptations, as well as enhanced anaerobic capacity of skeletal muscle (Škarabot et al., 2021; Zanuso et al., 2017), although the evidence of the effect of exercise on these outcomes in individuals with neuropathy remains to be explored.

Reduced pain sensation and electrophysiological dysfunction, accompanied by increases in neurotrophic factors, have been observed in response to resistance training, although to a lesser extent compared to aerobic training (Cefis et al., 2023; Dobson et al., 2014; Groover et al., 2013; Ma et al., 2019; Zhang et al., 2021). The physiological effects associated with neurotrophic factor secretions promote reversing myelinic damage and restoring Ca^{2+} ionic channels, explaining the enhanced neural input transmission (Shankarappa et al., 2011).

The role of resistance training on the NMJ is currently unknown. However, it has been reported that NMJ responses to resistance training primarily regard structural expansion, particularly in the post-synaptic density of higher-threshold motor units (Deschenes, 2019). These modifications may also be beneficial for diabetic individuals with respect to mitigating the detrimental effects of diabetes-related complications on neuromuscular transmission.

Neurogenic sarcopenia and muscle atrophy are critical in diabetes-related complications, which are primarily mitigated through resistance training (Zacker, 2005). Indeed, the overloading of muscle structures induces hypertrophy, enhanced body mass index, waist circumference and fat mass (Acosta-Manzano et al., 2020). Furthermore, resistance training induces musculoskeletal adaptations through the combined effect of mechanical overload (Roberts et al., 2023) and time under tension (Burd et al., 2012), resulting in the upregulation of mechanistic target of rapamycin complex 1 (mTORc1) and follistatin, which is responsible for inhibiting myostatin (Amthor et al., 2004; Rodino-Klapac et al., 2009).

Resistance training is also associated with the production of *exerkines*, which is responsible for tissue regeneration and remodelling, paramount for mitigating diabetes-related complications (Chow et al., 2022; Robbins & Gerszten, 2023). As a result, the increase in muscle strength, accompanied by enhanced muscle quality and functionality, directly depends on the protective and preventive action of exercise training. Nevertheless, it is crucial to note that resistance training adaptations are severely attenuated with ageing, although a significant protective effect has been recognised (Hepple & Rice, 2016).

Combined exercise training. The physiological mechanisms associated with the beneficial effects of combined aerobic and resistance training on diabetic individuals directly incorporate the abovementioned evidence of either isolated exercise modalities. However, combining these training modalities represents an additional strategy for optimising gains from both in diabetic individuals. Recent findings indicate that combined exercise training induces a significant reduction

in the body mass index of $\sim 0.98 \text{ kg m}^{-2}$, accompanied by lower body fat percentage and a preserved muscle mass (Zhao et al., 2021). The observed effects depend on the physiological response to mechanical overload in promoting hypertrophy and the higher metabolic rate promoted by aerobic exercise, further contributing to physical fitness, weight management and individual independence (Coffey & Hawley, 2017; Huiberts et al., 2024; Maiorana et al., 2002). Combined exercise also promotes significant reductions in both systolic and diastolic blood pressure and enhancements in lipid profiles in diabetic individuals (Zhao et al., 2021). These modifications contribute to reduced cardiovascular risk, improved vascular health and overall enhancements in blood lipid regulation in diabetic individuals, especially those with T2D, as a result of the close association with a sedentary lifestyle (Pinto et al., 2023).

The combined benefits of aerobic and resistance training increase confidence and satisfaction with physical capabilities, which is positively associated with exercise adherence in a relatively long-term intervention (Balducci et al., 2010). Improved adherence strengthens the likelihood of continued metabolic and functional benefits, supporting the long-term health of diabetic patients. However, mild musculoskeletal discomfort or the aggravation of pre-existing conditions, such as osteoarthritis, can occur, particularly in unsupervised settings (Zhao et al., 2021). Although these events are typically minor, proper supervision and individualised adjustments to exercise regimens are recommended to minimise risk, emphasising the importance of individualised, guided exercise programs for the diabetic population.

The role of physical exercise in glycaemic control. To mitigate the detrimental effects of glycaemic variability, 150–300 min of moderate-intensity or 75–150 min of vigorous-intensity aerobic exercise per week and at least two sessions per week of resistance exercise have been suggested in people with diabetes (Kanaley et al., 2022; Pinto et al., 2023). The aim is to minimise sedentary habits by performing activity breaks or any movement, based on the motto ‘every move counts’ (Bull et al., 2020). This is achieved through aerobic exercise, resistance training and combining both approaches (Coffey & Hawley, 2017; Kanaley et al., 2022; Maiorana et al., 2002).

Aerobic training, including continuous or interval-based exercise, is strongly associated with improved glycaemic control in both healthy and diabetic individuals. Regular aerobic exercise has been linked to reductions in HbA1c and fasting glucose levels, alongside enhanced insulin sensitivity and lipid management, which are particularly beneficial in the diabetic population (Ashcroft et al., 2024; Bellini et al., 2024; Bird & Hawley, 2017; Haxhi & Bellini, 2024; Van Dijk & Van Loon, 2015).

At the molecular level, aerobic exercise promotes glucose uptake in skeletal muscle primarily through the activation of AMP-activated protein kinase (AMPK), a pathway that facilitates the translocation of glucose transporter type 4 GLUT (GLUT4) to the cell membrane (Fueger et al., 2007; Kennedy et al., 1999; Leprivier & Rotblat, 2020). This insulin-independent mechanism of GLUT4 mobilisation is essential for improving glycaemic control, particularly in individuals with impaired insulin signalling. Although exercise does not appear to affect insulin receptor activity (Treadway et al., 1989), GLUT4 translocation has been suggested to depend on NO signalling and calmodulin-dependent protein kinase pathways associated with muscle contraction (Knudsen et al., 2020; Richter & Hargreaves, 2013; Sjøberg et al., 2017). Notably, this insulin receptor-independent action contributed to the recognition of physical exercise as the most potent stimulus to increase skeletal muscle GLUT4 expression, thereby improving glycaemic control and underscoring the protective role of exercise in diabetes-related complications.

Aerobic exercise also stimulates IL-6 release from muscle, which subsequently enhances glucagon-like peptide-1 secretion from intestinal L-cells and the pancreas, indirectly improving insulin response and glycaemic regulation (Holst, 2007; Morettini et al., 2017; Müller et al., 2019). This cascade reduces systemic inflammation, improves fat oxidation and enhances cardiovascular health, further supporting glucose homeostasis and long-term metabolic health in diabetic individuals (Karstoft & Pedersen, 2016).

Resistance training also enhances glycaemic control through different mechanisms from aerobic exercise. Unlike aerobic training, resistance exercise does not primarily rely on AMPK activation but rather on mTOR-related pathways, which are stimulated by the muscle contractions and microtears characteristic of strength exercises (Röhling et al., 2016). This mTOR pathway activation enhances glucose transport into skeletal muscle cells, improving insulin sensitivity and aiding in glycaemic control. Resistance training also enhances the release of myokines, comprising proteins that play a role in muscle signalling and metabolic regulation. Myokines released following resistance exercise modulate inflammation, promote fat oxidation and improve -cell function, helping to reduce endothelial dysfunction and mitigate cardiovascular risks associated with diabetes (Karstoft & Pedersen, 2016; Pedersen, 2017).

Even though exercise substantially improves life quality in diabetic individuals, many patients do not exercise regularly. One of the leading causes of poor exercise program adherence is fear of hypoglycaemic events during the session, especially among T1D individuals (Riddell et al., 2017). Hyper- and hypoglycaemic events during exercise are relevant issues that these patients have to

Table 2. Training strategies for diabetic individuals

| Training | Prescription | Adaptation | References |
|--|--|--|--|
| Endurance training continuous activity at moderate and high-intensity of running/swimming/cycling | 55–75%HRmax RPE 12–13 >150 min week ⁻¹ 3–7 days week ⁻¹ 75–95%HRmax RPE 14–16 75–150 min week ⁻¹ 3–7 days week ⁻¹ No more than 48 h of rest | HbA1c level ↓ Insulin sensitivity ↑ Blood pressure ↓ Lipid profile ↑ Fat mass ↓ Glycaemic control ↑ Mitochondrial density ↑ | (Colberg et al., 2016; Rodino-Klapac et al., 2009) |
| Strength training Resistance machines, free weights, resistance bands, and/or body weight as resistance exercises | 1–3 sets, 10–15 repetitions OMNI 7–8 8–10 exercises per session 1–3 sets, <8 repetitions OMNI 8–9 8–10 exercises per session Minimum 48 h of rest | HbA1C ↓↓ Fat mass ↓ Strength ↑↑ Sarcopenia ↓↓↓ Body composition ↑↑ | (Acosta-Manzano et al., 2020; Kanaley et al., 2022; Syeda et al., 2023) |
| HIIT High-intensity repeated bouts of running/swimming/cycling or resistance training | 10 s to 4 min, high-intensity (75–95%HRmax) 12 s to 5 min, active/passive rest (30–60%HRmax) 3 days week ⁻¹ for vigorous aerobic 2 days week ⁻¹ for resistance training | Glycaemic control ↑↑ Oxidative capacity ↑↑ Insulin sensitivity ↑↑ Stroke volume ↑ | (Van Ryckeghem et al., 2022; Wilson et al., 2019) |
| Combined training (strength + aerobic) resistance training (weight loaded) + aerobic exercises (i.e. walking, cycling) | Circuit training 7 laps, strength + aerobic Strength (60%1RM, 15 repetitions) Aerobic 70–80%HRmax 3 sessions week ⁻¹ 1 h per session | HbA1 ↓↓ Glycaemia ↓ Strength ↑ Waist: hip ratio ↓ Skinfold ↓↓ Body composition ↑↑ $\dot{V}_{O_2\max}$ ↑ | (Coffey & Hawley, 2017; Huiberts et al., 2024; Maiorana et al., 2002; X. Su et al., 2022) |

↑ Slight increase, ↑↑ moderate increase, ↑↑↑ high increase, ↓ slight decrease, ↓↓ moderate decrease, ↓↓↓ high decrease. Classification was made based on Colberg et al. (2016) and Syeda et al. (2023). The OMNI scale has been adapted according to Helms et al. (2016); Lagally & Robertson (2006); Naclerio et al. (2011); and Robertson et al. (2003). HbA1C, haemoglobin A1C; HIIT, high-intensity interval training; HRmax, maximum heart rate; RM, repetition maximum; RPE, rating of perceived exertion.

deal with. Technological advances and the most recent studies on exercise safety have widely helped reduce these events and increase glucose control during exercise. In particular, evidence suggests that aerobic exercise elicits a greater glucose reduction during and several hours after the session compared to resistance exercise in T1D (Yardley et al., 2012) and T2D individuals (Bellini et al., 2023, 2024). This has mainly been associated with more production of growth hormone in response to resistance exercise, which mitigates broad excursions of glucose levels (Yardley et al., 2012). Therefore, it is paramount to consider combined exercise strategies to guarantee blood glucose stability both acutely (Minnock et al., 2020) and chronically (Minnock et al., 2022). Although the importance of exercise in managing blood glucose levels in diabetic individuals has been broadly discussed,

evaluating its impact on neuropathic complications still requires more investigation, underlining the physiological relevance of the beneficial effects promoted by physical exercise.

Guidelines for exercise prescription in diabetic individuals are reported in Table 2.

Practical applications

Regular exercise is crucial for treating diabetes-associated complications and shall be considered as the leading approach for therapeutic benefits (McGee & Hargreaves, 2020). Aerobic exercise improves vascular health, enhancing blood flow and oxygen delivery to tissues, and also promotes the release of BDNF, which supports

nerve health and function. Resistance training helps prevent and manage sarcopenia by enhancing muscle strength and preserving fat-free mass. These combined effects lead to better glycaemic control, reduced neuropathic symptoms and improved overall physical health. The abovementioned information finds applications throughout various exercise training modalities that shall be individualised and applied based on patients' needs.

Conclusions

Diabetic neuropathy is associated with deterioration in both the structural and functional aspects of the neuromuscular system, manifesting through impairments at the central and peripheral nervous systems, defective neuromuscular junction and accelerated loss of muscle fibres. These complications contribute to a severe decline in performance and independence, burdening the affected population. Current knowledge highlights the significant impact of hyperglycaemia-induced oxidative stress, impaired nerve regeneration and vascular complications on neuromuscular integrity. Although the precise mechanisms, particularly regarding CNS involvement, remain partially understood, evidence supports the critical role of glycaemic control in mitigating these complications. Notably, regular physical exercise emerges as a promising intervention, potentially counteracting neuromuscular decline by promoting muscle strength, hypertrophy, metabolic health and nerve regeneration. Future research should aim to elucidate the detailed pathways of neuromuscular deterioration to optimise the exercise protocols and therapeutic benefits for diabetic individuals and also include motor unit assessment over a broad range of modalities to elucidate the mechanisms underlying the loss of functional properties.

References

- Aagaard, P., Suetta, C., Caserotti, P., Magnusson, S. P., & Kjær, M. (2010). Role of the nervous system in sarcopenia and muscle atrophy with aging: Strength training as a countermeasure. *Scandinavian Journal of Medicine and Science in Sports*, **20**(1), 49–64.
- Acosta-Manzano, P., Rodriguez-Ayllon, M., Acosta, F., Niederseer, D., & Niebauer, J. (2020). Beyond general resistance training. Hypertrophy versus muscular endurance training as therapeutic interventions in adults with type 2 diabetes mellitus: A systematic review and meta-analysis. *Obesity Reviews*, **21**(6), e13007.
- Allen, M. D., Choi, I. H., Kimpinski, K., Doherty, T. J., & Rice, C. L. (2013). Motor unit loss and weakness in association with diabetic neuropathy in humans. *Muscle and Nerve*, **48**(2), 298–300.
- Allen, M. D., Doherty, T. J., Rice, C. L., & Kimpinski, K. (2016). Physiology in medicine: Neuromuscular consequences of diabetic neuropathy. *Journal of Applied Physiology*, **121**(1), 1–6.
- Allen, M. D., Kimpinski, K., Doherty, T. J., & Rice, C. L. (2014). Length dependent loss of motor axons and altered motor unit properties in human diabetic polyneuropathy. *Clinical Neurophysiology*, **125**(4), 836–843.
- Allen, M. D., Kimpinski, K., Doherty, T. J., & Rice, C. L. (2015). Decreased muscle endurance associated with diabetic neuropathy may be attributed partially to neuromuscular transmission failure. *Journal of Applied Physiology*, **118**(8), 1014–1022.
- Allen, M. D., Major, B., Kimpinski, K., Doherty, T. J., & Rice, C. L. (2014). Skeletal muscle morphology and contractile function in relation to muscle denervation in diabetic neuropathy. *Journal of Applied Physiology*, **116**(5), 545–552.
- Amthor, H., Nicholas, G., McKinnell, I., Kemp, C. F., Sharma, M., Kambadur, R., & Patel, K. (2004). Follistatin complexes Myostatin and antagonises Myostatin-mediated inhibition of myogenesis. *Developmental Biology*, **270**(1), 19–30.
- Anagnostou, M., & Hepple, R. (2020). Mitochondrial mechanisms of neuromuscular junction degeneration with aging. *Cells*, **9**(1), 197.
- Andersen, H. (1999). Motor function in diabetic neuropathy. *Acta Neurologica Scandinavica*, **100**(4), 211–220.
- Andersen, H. (2012). Motor dysfunction in diabetes. *Diabetes/Metabolism Research and Reviews*, **28**(Suppl 1), 89–92.
- Andersen, H., Nielsen, S., Mogensen, C. E., & Jakobsen, J. (2004). Muscle strength in type 2 diabetes. *Diabetes*, **53**(6), 1543–1548.
- Andersen, H., Poulsen, P. L., Mogensen, C. E., & Jakobsen, J. (1996). Isokinetic muscle strength in long-term IDDM patients in relation to diabetic complications. *Diabetes*, **45**(4), 440–445.
- Andersen, H., Stålberg, E., Gjerstad, M. D., & Jakobsen, J. (1998). Association of muscle strength and electrophysiological measures of reinnervation in diabetic neuropathy. *Muscle and Nerve*, **21**(12), 1647–1654.
- Andreassen, C. S., Jakobsen, J., & Andersen, H. (2006). Muscle weakness: A progressive late complication in diabetic distal symmetric polyneuropathy. *Diabetes*, **55**(3), 806–812.
- Ashcroft, S. P., Stocks, B., Egan, B., & Zierath, J. R. (2024). Exercise induces tissue-specific adaptations to enhance cardiometabolic health. *Cell Metabolism*, **36**(2), 278–300.
- Assar, M. E.I., Laosa, O., & Mañas, L. R. (2019). Diabetes and frailty. *Current Opinion in Clinical Nutrition and Metabolic Care*, **22**(1), 52–57.
- Avaltroni, P., Cappellini, G., Sylos-Labini, F., Ivanenko, Y., & Lacquaniti, F. (2024). Spinal maps of motoneuron activity during human locomotion: Neuromechanical considerations. *Frontiers in Physiology*, **15**, 1389436.
- Bacchi, E., Negri, C., Zanolin, M. E., Milanese, C., Faccioli, N., Trombetta, M., Zoppini, G., Cevese, A., Bonadonna, R. C., Schena, F., Bonora, E., Lanza, M., & Moghetti, P. (2012). Metabolic effects of aerobic training and resistance training in type 2 diabetic subjects: A randomized controlled trial (the RAED2 study). *Diabetes Care*, **35**(4), 676–682.
- Badrah, M. H., Abdelaaty, T. A., Imbaby, S. A. E., Abdel-Fattah, Y. H., Silim, W. M., & El Feky, A. Y. (2022). The relationship between vascular endothelial growth factor-A serum level and the severity of diabetic peripheral neuropathy. *Egyptian Rheumatology and Rehabilitation*, **49**(1), 247–252.

- Bahadoran, Z., Mirmiran, P., Kashfi, K., & Ghasemi, A. (2023). Vascular nitric oxide resistance in type 2 diabetes. In *Cell Death and Disease* (Vol. 14, Issue 7). Springer Nature.
- Balducci, S., Iacobellis, G., Parisi, L., Di Biase, N., Calandriello, E., Leonetti, F., & Fallucca, F. (2006). Exercise training can modify the natural history of diabetic peripheral neuropathy. *Journal of Diabetes and Its Complications*, **20**(4), 216–223.
- Balducci, S., Zanuso, S., Nicolucci, A., Fernando, F., Cavallo, S., Cardelli, P., Fallucca, S., Alessi, E., Letizia, C., Jimenez, A., Fallucca, F., & Pugliese, G. (2010). Anti-inflammatory effect of exercise training in subjects with type 2 diabetes and the metabolic syndrome is dependent on exercise modalities and independent of weight loss. *Nutrition, Metabolism and Cardiovascular Diseases*, **20**(8), 608–617.
- Ballabh, P., Braun, A., & Nedergaard, M. (2004). The blood-brain barrier: An overview: Structure, regulation, and clinical implications. *Neurobiology of Disease*, **16**(1), 1–13.
- Bannister, K. (2019). Descending pain modulation: Influence and impact. *Current Opinion in Physiology*, **11**, 62–66.
- Bannister, K., & Dickenson, A. H. (2017). The plasticity of descending controls in pain: Translational probing. *The Journal of Physiology*, **595**(13), 4159–4166.
- Barakou, I., Sakalidis, K. E., Abonie, U. S., Finch, T., Hackett, K. L., & Hettinga, F. J. (2023). Effectiveness of physical activity interventions on reducing perceived fatigue among adults with chronic conditions: A systematic review and meta-analysis of randomised controlled trials. *Scientific Reports*, **13**(1), 14582.
- Basu, R., Johnson, M. L., Kudva, Y. C., & Basu, A. (2014). Exercise, hypoglycemia, and type 1 diabetes. In *Diabetes Technology and Therapeutics* (Vol. 16, Issue 6, pp. 331–337). Mary Ann Liebert Inc.
- Baynes, J. W. (1991). Role of oxidative stress in development of complications in diabetes. *Diabetes*, **40**(4), 405–412.
- Bazzucchi, I., De Vito, G., Felici, F., Dewhurst, S., Sgadari, A., & Sacchetti, M. (2015). Effect of exercise training on neuromuscular function of elbow flexors and knee extensors of type 2 diabetic patients. *Journal of Electromyography and Kinesiology*, **25**(5), 815–823.
- Behrens, M., Gube, M., Chaabene, H., Prieske, O., Zenon, A., Broscheid, K. C., Schega, L., Husmann, F., & Weippert, M. (2023). Fatigue and human performance: An updated framework. *Sports Medicine*, **53**(1), 7–31.
- Beigi, S., Shabkhiz, F., Kordi, M., Haghi-Ashtiani, B., Hashemi-Madani, N., & Zmijewski, P. (2023). The effects of a 10-week aerobic and unilateral lower extremity resistance training program on amplitude and nerve conduction velocity of sensory and motor nerves in diabetic patients with neuropathy. *Journal of Human Kinetics*, **87**, 93–103.
- Bellini, A., Nicolò, A., Bulzomi, R., Bazzucchi, I., & Sacchetti, M. (2021). The effect of different postprandial exercise types on glucose response to breakfast in individuals with type 2 diabetes. *Nutrients*, **13**(5), 1440.
- Bellini, A., Nicolò, A., Rocchi, J. E., Bazzucchi, I., & Sacchetti, M. (2023). Walking attenuates postprandial glycemic response: What else can we do without leaving home or the office? *International Journal of Environmental Research and Public Health*, **20**(1), 253.
- Bellini, A., Scotto di Palumbo, A., Nicolò, A., Bazzucchi, I., & Sacchetti, M. (2024). Exercise prescription for postprandial glycemic management. *Nutrients*, **16**(8), 1170.
- Bensellam, M., Laybutt, D. R., & Jonas, J. C. (2012). The molecular mechanisms of pancreatic β -cell glucotoxicity: Recent findings and future research directions. *Molecular and Cellular Endocrinology*, **364**(1–2), 1–27.
- Bilchak, J. N., Caron, G., & Côté, M. P. (2021). Exercise-induced plasticity in signaling pathways involved in motor recovery after spinal cord injury. *International Journal of Molecular Sciences*, **22**(9), 4858.
- Bird, S. R., & Hawley, J. A. (2017). Update on the effects of physical activity on insulin sensitivity in humans. *British Medical Journal Open Sport and Exercise Medicine*, **2**(1), e000143.
- Blair, N. T., & Bean, B. P. (2002). Roles of tetrodotoxin (TTX)-sensitive Na^+ current, TTX-resistant Na^+ current, and Ca^{2+} current in the action potentials of nociceptive sensory neurons. *Journal of Neuroscience*, **22**(23), 10277–10290.
- Boulé, N. G., Kenny, G. P., Haddad, E., Wells, G. A., & Sigal, R. J. (2003). Meta-analysis of the effect of structured exercise training on cardiorespiratory fitness in Type 2 diabetes mellitus. *Diabetologia*, **46**(8), 1071–1081.
- Bull, F. C., Al-Ansari, S. S., Biddle, S., Borodulin, K., Buman, M. P., Cardon, G., Carty, C., Chaput, J. P., Chastin, S., Chou, R., Dempsey, P. C., Dipietro, L., Ekelund, U., Firth, J., Friedenreich, C. M., Garcia, L., Gichu, M., Jago, R., Katzmarzyk, P. T., ... Willumsen, J. F. (2020). World Health Organization 2020 guidelines on physical activity and sedentary behaviour. In *British Journal of Sports Medicine* (Vol. 54, Issue 24, pp. 1451–1462). BMJ Publishing Group.
- Burd, N. A., Andrews, R. J., West, D. W. D., Little, J. P., Cochran, A. J. R., Hector, A. J., Cashaback, J. G. A., Gibala, M. J., Potvin, J. R., Baker, S. K., & Phillips, S. M. (2012). Muscle time under tension during resistance exercise stimulates differential muscle protein sub-fractional synthetic responses in men. *The Journal of Physiology*, **590**(2), 351–362.
- Calderón, J. C., Bolaños, P., & Caputo, C. (2014). The excitation-contraction coupling mechanism in skeletal muscle. *Biophysical Reviews*, **6**(1), 133–160.
- Callaghan, B., & Feldman, E. (2013). The metabolic syndrome and neuropathy: Therapeutic challenges and opportunities. *Annals of Neurology*, **74**(3), 397–403.
- Cefis, M., Chaney, R., Wirtz, J., Méloux, A., Quirié, A., Leger, C., Prigent-Tessier, A., & Garnier, P. (2023). Molecular mechanisms underlying physical exercise-induced brain BDNF overproduction. *Frontiers in Molecular Neuroscience*, **16**, 1275924.
- Ceriello, A., Esposito, K., Inthan, M., Thorpe, J., & Giuliano, D. (2010). Effect of acute hyperglycaemia, long-term glycaemic control and insulin on endothelial dysfunction and inflammation in Type 1 diabetic patients with different characteristics. *Diabetic Medicine*, **27**(8), 911–917.
- Ceriello, A., & Ihnat, M. (2010). ‘Glycaemic variability’: A new therapeutic challenge in diabetes and the critical care setting. *Diabetic Medicine*, **27**(8), 862–867.

- Ceriello, A., Quagliaro, L., Piconi, L., Assaloni, R., Da Ros, R., Maier, A., Esposito, K., & Giugliano, D. (2004). *Effect of postprandial hypertriglyceridemia and hyperglycemia on circulating adhesion molecules and oxidative stress generation and the possible role of simvastatin treatment*. <http://diabetesjournals.org/diabetes/article-pdf/53/3/701/656831/zdb00304000701.pdf>
- Cernea, S., & Raz, I. (2021). Management of diabetic neuropathy. *Metabolism: Clinical and Experimental*, **123**, 154867.
- Chang, M. C., Park, C. R., Rhie, S. H., Shim, W. H., & Kim, D. Y. (2019). Early treadmill exercise increases macrophage migration inhibitory factor expression after cerebral ischemia/reperfusion. *Neural Regeneration Research*, **14**(7), 1230–1236.
- Chavanelle, V., Boisseau, N., Otero, Y. F., Combaret, L., Darvet, D., Montaurier, C., Delcros, G., Peltier, S. L., & Sirvent, P. (2017). Effects of high-intensity interval training and moderate-intensity continuous training on glycaemic control and skeletal muscle mitochondrial function in db/db mice. *Scientific Reports*, **7**(1), 204.
- Chen, J., Zhou, R., Feng, Y., & Cheng, L. (2022). Molecular mechanisms of exercise contributing to tissue regeneration. *Signal Transduction and Targeted Therapy*, **7**(1), 383.
- Choi, S. E., & Rush, E. B. (2012). Effect of a short-duration, culturally tailored, community-based diabetes self-management intervention for Korean immigrants: A pilot study. *The Diabetes Educator*, **38**(3), 377–385.
- Chow, L. S., Gerszten, R. E., Taylor, J. M., Pedersen, B. K., van Praag, H., Trappe, S., Febbraio, M. A., Galis, Z. S., Gao, Y., Haus, J. M., Lanza, I. R., Lavie, C. J., Lee, C. H., Lucia, A., Moro, C., Pandey, A., Robbins, J. M., Stanford, K. I., Thackray, A. E., ... Snyder, M. P. (2022). Exerkines in health, resilience and disease. *Nature Reviews Endocrinology*, **18**(5), 273–289.
- Cigrovski Berkovic, M., Bilic-Curcic, I., La Grasta Sabolic, L., Mrzljak, A., & Cigrovski, V. (2021). Fear of hypoglycemia, a game changer during physical activity in type 1 diabetes mellitus patients. *World Journal of Diabetes*, **12**(5), 569–577.
- Coffey, V. G., & Hawley, J. A. (2017). Concurrent exercise training: Do opposites distract? In *Journal of Physiology* (Vol., **595**, Issue 9, pp. 2883–2896). Blackwell Publishing Ltd.
- Colberg, S. R., Sigal, R. J., Yardley, J. E., Riddell, M. C., Dunstan, D. W., Dempsey, P. C., Horton, E. S., Castorino, K., & Tate, D. F. (2016). Physical activity/exercise and diabetes: A position statement of the American Diabetes Association. In *Diabetes Care* (Vol., **39**, Issue 11, pp. 2065–2079). American Diabetes Association Inc.
- Coppey, L. J., Davidson, E. P., Dunlap, J. A., Lund, D. D., & Yorek, M. A. (2000). Slowing of motor nerve conduction velocity in streptozotocin-induced diabetic rats is preceded by impaired vasodilation in arterioles that overlie the sciatic nerve. *Experimental Diabetes Research*, **1**(2), 131–143.
- D'Antona, G., Pellegrino, M. A., Adami, R., Rossi, R., Naccari Carlizzi, C., Canepari, M., Saltin, B., & Bottinelli, R. (2003). The effect of ageing and immobilization on structure and function of human skeletal muscle fibres. *The Journal of Physiology*, **552**(2), 499–511.
- de Oliveira, V. N., Bessa, A., Jorge, M. L. M. P., da Silva Oliveira, R. J., de Mello, M. T., de Agostini, G. G., Jorge, P. T., & Espindola, F. S. (2012). The effect of different training programs on antioxidant status, oxidative stress, and metabolic control in type 2 diabetes. *Applied Physiology, Nutrition and Metabolism*, **37**(2), 334–344.
- del Vecchio, A., Enoka, R. M., & Farina, D. (2024). Specificity of early motor unit adaptations with resistive exercise training. *The Journal of Physiology*, **602**(12), 2679–2688.
- del Vecchio, A., Negro, F., Holobar, A., Casolo, A., Folland, J. P., Felici, F., & Farina, D. (2019). You are as fast as your motor neurons: Speed of recruitment and maximal discharge of motor neurons determine the maximal rate of force development in humans. *The Journal of Physiology*, **597**(9), 2445–2456.
- Delevatti, R. S., Bracht, C. G., Lisboa, S. D. C., Costa, R. R., Marson, E. C., Netto, N., & Kruehl, L. F. M. (2019). The role of aerobic training variables progression on glycemic control of patients with type 2 diabetes: A systematic review with meta-analysis. *Sports Medicine - Open*, **5**(1), 22.
- Deschenes, M. R. (2019). Adaptations of the neuromuscular junction to exercise training. *Current Opinion in Physiology*, **10**, 10–16.
- Dixit, S., Maiya, A., & Shastry, B. (2014). Effect of aerobic exercise on quality of life in population with diabetic peripheral neuropathy in type 2 diabetes: A single blind, randomized controlled trial. *Quality of Life Research*, **23**(5), 1629–1640.
- Dobson, J. L., McMillan, J., & Li, L. (2014). Benefits of exercise intervention in reducing neuropathic pain. In *Frontiers in Cellular Neuroscience* (Vol., **8**, Issue 1 APR). Frontiers Research Foundation.
- D'Souza, D. M., Al-Sajee, D., & Hawke, T. J. (2013). Diabetic myopathy: Impact of diabetes mellitus on skeletal muscle progenitor cells. *Frontiers in Physiology*, **4**, 379.
- Dubin, A. E., & Patapoutian, A. (2010). Nociceptors: The sensors of the pain pathway. *Journal of Clinical Investigation*, **120**(11), 3760–3772.
- Duchateau, J., & Enoka, R. M. (2011). Human motor unit recordings: Origins and insight into the integrated motor system. *Brain Research*, **1409**, 42–61.
- English, A. W., Wilhelm, J. C., & Ward, P. J. (2014). Exercise, neurotrophins, and axon regeneration in the PNS. *Physiology*, **29**(6), 437–445.
- Enoka, R. M., & Fuglevand, A. J. (2001). Motor unit physiology: Some unresolved issues. *Muscle and Nerve*, **24**(1), 4–17.
- Estrada-Bonilla, Y. C., Castro, P. A. T. S., Luna, G. L. F., Souza, A. B. A., Santos, G. S., Salvini, T. F., Leal, A. M. O., & Russo, T. L. (2020). Reaching task performance is associated to neuromuscular junction adaptations in rats with induced diabetes mellitus. *Brazilian Journal of Medical and Biological Research*, **53**(7), e8763.
- Fahim, M. A., El-Sabban, F., & Davidson, N. (1998). Muscle contractility decrement and correlated morphology during the pathogenesis of streptozotocin-diabetic mice. *Anatomical Record*, **251**(2), 240–244.

- Fahim, M. A., Hasan, M. Y., & Alshuaib, W. B. (2000). Early morphological remodeling of neuromuscular junction in a murine model of diabetes. *Journal of Applied Physiology*, **89**(6), 2235–2240.
- Fang, F., Luo, Q., Ge, R. B., Lai, M. Y., Gong, Y. J., Kang, M., Ma, M. M., Zhang, L., Li, Y., Wang, Y. F., & Peng, Y. D. (2021). Decreased microstructural integrity of the central somatosensory tracts in diabetic peripheral neuropathy. *Journal of Clinical Endocrinology and Metabolism*, **106**(6), 1566–1575.
- Farina, D., & Gandevia, S. (2024). The neural control of movement: A century of in vivo motor unit recordings is the legacy of Adrian and Bronk. *The Journal of Physiology*, **602**(2), 281–295.
- Favretto, M. A., Andreis, F. R., Cossul, S., Negro, F., Oliveira, A. S., & Marques, J. L. B. (2023). Differences in motor unit behavior during isometric contractions in patients with diabetic peripheral neuropathy at various disease severities. *Journal of Electromyography and Kinesiology*, **68**, 102725.
- Favretto, M. A., Cossul, S., Andreis, F. R., & Marques, J. L. B. (2019). Evaluation of rate of muscular force development in type 2 diabetic individuals with and without diabetic peripheral neuropathy. *International Federation for Medical and Biological Engineering Proceedings*, **70**(1), 31–36.
- Feldman, E. L., Callaghan, B. C., Pop-Busui, R., Zochodne, D. W., Wright, D. E., Bennett, D. L., Brill, V., Russell, J. W., & Viswanathan, V. (2019). Diabetic neuropathy. In *Nature Reviews Disease Primers* (Vol. 5, Issue 1). Nature Publishing Group.
- Feldman, E. L., Nave, K. A., Jensen, T. S., & Bennett, D. L. H. (2017). New horizons in diabetic neuropathy: Mechanisms, bioenergetics, and pain. *Neuron*, **93**(6), 1296–1313.
- Forbes, J. M., & Cooper, M. E. (2013). Mechanisms of diabetic complications. *Physiological Reviews*, **93**(1), 137–188.
- Francis, G. J., Martinez, J. A., Liu, W. Q., Zochodne, D. W., Hanson, L. R., Frey, W. H., & Toth, C. (2011). Motor end plate innervation loss in diabetes and the role of insulin. *Journal of Neuropathology and Experimental Neurology*, **70**(5), 323–339.
- Fritschi, C., & Quinn, L. (2010). Fatigue in patients with diabetes: A review. *Journal of Psychosomatic Research*, **69**(1), 33–41.
- Fueger, P. T., Li, C. Y., Ayala, J. E., Shearer, J., Bracy, D. P., Charron, M. J., Rottman, J. N., & Wasserman, D. H. (2007). Glucose kinetics and exercise tolerance in mice lacking the GLUT4 glucose transporter. *The Journal of Physiology*, **582**(2), 801–812.
- Gandevia, S. C. (2001). Spinal and supraspinal factors in human muscle fatigue. *Physiological Reviews*, **81**(4), 1725–1789.
- Gandevia, S. C., Macefield, G., Burke, D., & McKenzie, D. K. (1990). Voluntary activation of human motor axons in the absence of muscle afferent feedback: The control of the deafferented hand. *Brain*, **113**(5), 1563–1581.
- Garcia, C. C., Potian, J. G., Hognason, K., Thyagarajan, B., Sultatos, L. G., Souayah, N., Routh, V. H., & McArdle, J. J. (2012). Acetylcholinesterase deficiency contributes to neuromuscular junction dysfunction in type 1 diabetic neuropathy. *American Journal of Physiology - Endocrinology and Metabolism*, **303**(4), E551–E561.
- Gholami, F., Nikookheslat, S., Salekzamani, Y., Boule, N., & Jafari, A. (2018). Effect of aerobic training on nerve conduction in men with type 2 diabetes and peripheral neuropathy: A randomized controlled trial. *Neurophysiologie Clinique*, **48**(4), 195–202.
- Giacco, F., & Brownlee, M. (2010). Oxidative stress and diabetic complications. *Circulation Research*, **107**(9), 1058–1070.
- Giannopoulou, I., Fernhall, B., Carhart, R., Weinstock, R. S., Baynard, T., Figueroa, A., & Kanaley, J. A. (2005). Effects of diet and/or exercise on the adipocytokine and inflammatory cytokine levels of postmenopausal women with type 2 diabetes. *Metabolism: Clinical and Experimental*, **54**(7), 866–875.
- Giugliano, D., Ceriello, A., & Paolisso, G. (1996). Oxidative stress and diabetic vascular complications. *Diabetes Care*, **19**(3), 257–267.
- Goulding, R. P., Roche, D. M., Scott, S. N., Koga, S., Weston, P. J., & Marwood, S. (2020). Limitations to exercise tolerance in type 1 diabetes: The role of pulmonary oxygen uptake kinetics and priming exercise. *Journal of Applied Physiology*, **128**(5), 1299–1309.
- Groen, B. B. L., Hamer, H. M., Snijders, T., Van Kranenburg, J., Frijns, D., Vink, H., & Van Loon, L. J. C. (2014). Skeletal muscle capillary density and microvascular function are compromised with aging and type 2 diabetes. *Journal of Applied Physiology*, **116**(8). <https://doi.org/10.1152/jappphysiol.00919.2013>
- Groover, A. L., Ryals, J. M., Guilford, B. L., Wilson, N. M., Christianson, J. A., & Wright, D. E. (2013). Exercise-mediated improvements in painful neuropathy associated with prediabetes in mice. *Pain*, **154**(12), 2658–2667.
- Gu, Y., Qiu, Z. L., Liu, D. Z., Sun, G. L., Guan, Y. C., Hei, Z. Q., & Li, X. (2018). Differential gene expression profiling of the sciatic nerve in type 1 and type 2 diabetic mice. *Biomedical Reports*, **9**(4), 291–304.
- Haines, M. S., Leong, A., Porneala, B. C., Meigs, J. B., & Miller, K. K. (2022). Association between muscle mass and diabetes prevalence independent of body fat distribution in adults under 50 years old. *Nutrition and Diabetes*, **12**(1), 29.
- Hajer, G. R., van Haeften, T. W., & Visseren, F. L. J. (2008). Adipose tissue dysfunction in obesity, diabetes, and vascular diseases. *European Heart Journal*, **29**(24), 2959–2971.
- Halling, J. F., & Pilegaard, H. (2020). PGC-1 α -mediated regulation of mitochondrial function and physiological implications. *Applied Physiology, Nutrition and Metabolism*, **45**(9), 927–936.
- Hamid, W. S., Ahmed, H. S., Osman, M. A., & Babiker, R. (2021). Nerve conduction and its correlations with duration of diabetes mellitus and glycosylated haemoglobin in type 2 diabetes mellitus (T2DM). *Journal of Endocrinology, Metabolism and Diabetes of South Africa*, **26**(2), 46–51.
- Haque, F., Reaz, M. B. I., Ali, S. H. M., Arsad, N., & Chowdhury, M. E. H. (2020). Performance analysis of non-invasive electrophysiological methods for the assessment of diabetic sensorimotor polyneuropathy in clinical research: A systematic review and meta-analysis with trial sequential analysis. *Scientific Reports*, **10**(1), 21770.

- Harding, J. L., Pavkov, M. E., Magliano, D. J., Shaw, J. E., & Gregg, E. W. (2019). Global trends in diabetes complications: A review of current evidence. *Diabetologia*, **62**(1), 3–16.
- Haxhi, J., & Bellini, A. (2024). Esercizio fisico nel diabete mellito di tipo 1. *L'Endocrinologo*, **62**, 436–442.
- Heckman, C. J., & Enoka, R. M. (2012). Motor unit. *Comprehensive Physiology*, **2**(4), 2629–2682.
- Helms, E. R., Cronin, J., Storey, A., & Zourdos, M. C. (2016). Application of the repetitions in reserve-based rating of perceived exertion scale for resistance training. *Strength & Conditioning Journal*, **38**(4), 42–49.
- Henneman, E. (1957). Relation between size of neurons and their susceptibility to discharge. *Science*, **126**(3287), 1345–1347.
- Hepple, R. T., & Rice, C. L. (2016). Innervation and neuromuscular control in ageing skeletal muscle. In *Journal of Physiology* (Vol., **594**, Issue 8, pp. 1965–1978). Blackwell Publishing Ltd.
- Hernández, E. O., & Camilo Vanegas, O. (2015). Diabetic myopathy and mechanisms of disease. *Biochemistry & Pharmacology: Open Access*, **04**(05), e179.
- Heroux, M. E., Butler, A. A., Robertson, L. S., Fisher, G., & Gandevia, S. C. (2022). Proprioception: A new look at an old concept. *Journal of Applied Physiology*, **132**(3), 811–814.
- Hicks, C. W., & Selvin, E. (2019). Epidemiology of peripheral neuropathy and lower extremity disease in diabetes. *Current Diabetes Reports*, **19**(10), 86.
- Hilton, T. N., Tuttle, L. J., Bohnert, K. L., Mueller, M. J., & Sinacore, D. R. (2008). Excessive adipose tissue infiltration in skeletal muscle in individuals with obesity, diabetes mellitus, and peripheral neuropathy: Association with performance and function. *Physical Therapy*, **88**(11), 1336–1344.
- Ho, N., Sommers, M. S., & Lucki, I. (2013). Effects of diabetes on hippocampal neurogenesis: Links to cognition and depression. *Neuroscience and Biobehavioral Reviews*, **37**(8), 1346–1362.
- Holst, J. J. (2007). The physiology of glucagon-like peptide 1. *Physiological Reviews*, **87**(4), 1409–1439.
- Huebschmann, A. G., Kohrt, W. M., Herlache, L., Wolfe, P., Daugherty, S., Reusch, J. E., Bauer, T. A., & Regensteiner, J. G. (2015). Type 2 diabetes exaggerates exercise effort and impairs exercise performance in older women. *British Medical Journal Open Diabetes Research & Care*, **3**(1), e000124.
- Hughes, T. M., Ryan, C. M., Aizenstein, H. J., Nunley, K., Gianaros, P. J., Miller, R., Costacou, T., Strotmeyer, E. S., Orchard, T. J., & Rosano, C. (2013). Frontal gray matter atrophy in middle aged adults with type 1 diabetes is independent of cardiovascular risk factors and diabetes complications. *Journal of Diabetes and Its Complications*, **27**(6), 558–564.
- Huiberts, R. O., Wüst, R. C. I., & van der Zwaard, S. (2024). Concurrent strength and endurance training: A systematic review and meta-analysis on the impact of sex and training status. In *Sports Medicine* (Vol., **54**, Issue 2, pp. 485–503). Springer Science and Business Media Deutschland GmbH.
- Hur, J., O'Brien, P. D., Nair, V., Hinder, L. M., McGregor, B. A., Jagadish, H. V., Kretzler, M., Brosius, F. C., & Feldman, E. L. (2016). Transcriptional networks of murine diabetic peripheral neuropathy and nephropathy: Common and distinct gene expression patterns. *Diabetologia*, **59**(6), 1297–1306.
- Jagodac, M. M., Pathirathna, S., Nelson, M. T., Mancuso, S., Joksovic, P. M., Rosenberg, E. R., Bayliss, D. A., Jevtovic-Todorovic, V., & Todorovic, S. M. (2007). Cell-specific alterations of T-type calcium current in painful diabetic neuropathy enhance excitability of sensory neurons. *Journal of Neuroscience*, **27**(12), 3305–3316.
- Jingjing, T., Fan, J., & Zhang, T. (2023). Mitochondria as a target for exercise-mitigated type 2 diabetes. *Journal of Molecular Histology*, **54**(6), 543–557.
- Kalra, S., & Sahay, R. (2018). Diabetes fatigue syndrome. *Diabetes Therapy*, **9**(4), 1421–1429.
- Kalyani, R. R., Metter, E. J., Egan, J., Golden, S. H., & Ferrucci, L. (2015). Hyperglycemia predicts persistently lower muscle strength with aging. *Diabetes Care*, **38**(1), 82–90.
- Kanaley, J. A., Colberg, S. R., Corcoran, M. H., Malin, S. K., Rodriguez, N. R., Crespo, C. J., Kirwan, J. P., & Zierath, J. R. (2022). Exercise/physical activity in individuals with type 2 diabetes: A consensus statement from the American college of sports medicine. *Medicine and Science in Sports and Exercise*, **54**(2), 353–368.
- Karstoft, K., & Pedersen, B. K. (2016). Exercise and type 2 diabetes: Focus on metabolism and inflammation. In *Immunology and Cell Biology* (Vol., **94**, Issue 2, pp. 146–150). Nature Publishing Group.
- Kennedy, J. W., Hirshman, M. F., Gervino, E. V., Ocel, J. V., Forse, R. A., Hoenig, S. J., Aronson, D., Goodyear, L. J., & Horton, E. S. (1999). Acute exercise induces GLUT4 translocation in skeletal muscle of normal human subjects and subjects with type 2 diabetes. *Diabetes*, **48**(5), 1192–1197.
- Kimura, I., Okazaki, M., & Kimura, M. (1993). Streptozocin-diabetes modifies acetylcholine release from mouse phrenic nerve terminal and presynaptic sensitivity to succinylcholine. *The Japanese Journal of Pharmacology*, **62**(1), 35–41.
- Knudsen, J. R., Steenberg, D. E., Hingst, J. R., Hodgson, L. R., Henriquez-Olguin, C., Li, Z., Kiens, B., Richter, E. A., Wojtaszewski, J. F. P., Verkade, P., & Jensen, T. E. (2020). Prior exercise in humans redistributes intramuscular GLUT4 and enhances insulin-stimulated sarcolemmal and endosomal GLUT4 translocation. *Molecular Metabolism*, **39**, 100998.
- Labally, K. M., & Robertson, R. J. (2006). Construct validity of the OMNI resistance exercise scale. *Journal of Strength and Conditioning Research*, **20**(2), 252–256.
- Lawrence, D. G., & Locke, S. (1961). Motor nerve conduction velocity in diabetes. *Archives of Neurology*, **5**(5). <https://doi.org/10.1001/archneur.1961.00450170021003>
- Le Corre, A., Caron, N., Turpin, N. A., & Dalleau, G. (2023). Mechanisms underlying altered neuromuscular function in people with DPN. In *European Journal of Applied Physiology* (Vol., **123**, Issue 7, pp. 1433–1446). Springer Science and Business Media Deutschland GmbH.

- Leitzelar, B. N., & Koltyn, K. F. (2021). Exercise and neuropathic pain: A general overview of preclinical and clinical research. *Sports Medicine - Open*, **7**(1), 21.
- Leprivier, G., & Rotblat, B. (2020). How does mTOR sense glucose starvation? AMPK is the usual suspect. *Cell Death Discovery*, **6**(1), 27.
- Lesniewski, L. A., Miller, T. A., & Armstrong, R. B. (2003). Mechanisms of force loss in diabetic mouse skeletal muscle. *Muscle and Nerve*, **28**(4), 493–500.
- Li, L., Yang, Y., Bai, J., Zhang, Y., Yang, H., Zhang, Y., & Lv, H. (2022). Impaired vascular endothelial function is associated with peripheral neuropathy in patients with type 2 diabetes. *Diabetes, Metabolic Syndrome and Obesity*, **15**, 1437–1449.
- Li, S., Yuan, S., Zhang, J., Xu, F., & Zhu, F. (2024). The effect of periodic resistance training on obese patients with type 2 diabetic nephropathy. *Scientific Reports*, **14**(1), 2761.
- Li, Y., Liu, Y., Liu, S., Gao, M., Wang, W., Chen, K., Huang, L., & Liu, Y. (2023). Diabetic vascular diseases: Molecular mechanisms and therapeutic strategies. *Signal Transduction and Targeted Therapy*, **8**(1), 152.
- Liddell, E. G. T., & Sherrington, C. S. (1925). Recruitment and some other factors of reflex inhibition. *Proceedings of the Royal Society of London*, **B97**, 488–518.
- Lin, J., Zhang, X., Sun, Y., Xu, H., Li, N., Wang, Y., Tian, X., Zhao, C., Wang, B., Zhu, B., & Zhao, R. (2024). Exercise ameliorates muscular excessive mitochondrial fission, insulin resistance and inflammation in diabetic rats via irisin/AMPK activation. *Scientific Reports*, **14**(1), 19174.
- Lundby, C., & Jacobs, R. A. (2016). Adaptations of skeletal muscle mitochondria to exercise training. *Experimental Physiology*, **101**(1), 17–22.
- Luo, J., Zhu, H. Q., Gou, B., & Zheng, Y. L. (2022). Mechanisms of exercise for diabetic neuropathic pain. *Frontiers in Aging Neuroscience*, **14**, 975453.
- Ma, X. Q., Qin, J., Li, H. Y., Yan, X. L., Zhao, Y., & Zhang, L. J. (2019). Role of exercise activity in alleviating neuropathic pain in diabetes via inhibition of the pro-inflammatory signal pathway. *Biological Research for Nursing*, **21**(1), 14–21.
- Maffiuletti, N. A., Aagaard, P., Blazevich, A. J., Folland, J., Tillin, N., & Duchateau, J. (2016). Rate of force development: Physiological and methodological considerations. In *European Journal of Applied Physiology* (Vol. **116**, Issue 6, pp. 1091–1116). Springer Verlag.
- Maiorana, A., O'driscoll, G., Goodman, C., Taylor, R., & Green, D. (2002). Combined aerobic and resistance exercise improves glycemic control and fitness in type 2 diabetes. *Diabetes Research and Clinical Practice*, **56**(2), 115–123.
- Malone, J. I. (2016). Diabetic central neuropathy: CNS damage related to hyperglycemia. *Diabetes*, **65**(2), 355–357.
- Marques, M. J., & Neto, H. S. (2002). Acetylcholine receptors and nerve terminal distribution at the neuromuscular junction of non-obese diabetic mice. *Anatomical Record*, **267**(2), 112–119.
- Martinez-Valdes, E., Enoka, R. M., Holobar, A., McGill, K., Farina, D., Besomi, M., Hug, F., Falla, D., Carson, R. G., Clancy, E. A., Disselhorst-Klug, C., van Dieën, J. H., Tucker, K., Gandevia, S., Lowery, M., Søgaard, K., Besier, T., Merletti, R., Kiernan, M. C., ... Hodges, P. W. (2023). Consensus for experimental design in electromyography (CEDE) project: Single motor unit matrix. *Journal of Electromyography and Kinesiology*, **68**, 102726.
- McDonnell, M. N., Buckley, J. D., Opie, G. M., Ridding, M. C., & Semmler, J. G. (2013). A single bout of aerobic exercise promotes motor cortical neuroplasticity. *Journal of Applied Physiology*, **114**(9), 1174–1182.
- McGee, S., & Hargreaves, M. (2020). Exercise adaptations: Molecular mechanisms and potential targets for therapeutic benefit. *Nature Reviews Endocrinology*, **16**(9), 495–505.
- McGregor, C. E., & English, A. W. (2019). The role of BDNF in peripheral nerve regeneration: Activity-dependent treatments and Val66Met. *Frontiers in Cellular Neuroscience*, **12**, 522.
- McLean, D. L., Fan, J., Higashijima, S. I., Hale, M. E., & Fetcho, J. R. (2007). A topographic map of recruitment in spinal cord. *Nature*, **446**(7131), 71–75.
- Meacham, K., Shepherd, A., Mohapatra, D. P., & Haroutounian, S. (2017). Neuropathic pain: Central vs. peripheral mechanisms. *Current Pain and Headache Reports*, **21**(6), 1–11.
- Meex, R. C. R., Schrauwen-Hinderling, V. B., Moonen-Kornips, E., Schaart, G., Mensink, M., Phielix, E., van de Weijer, T., Sels, J. P., Schrauwen, P., & Hesselink, M. K. C. (2010). Restoration of muscle mitochondrial function and metabolic flexibility in type 2 diabetes by exercise training is paralleled by increased myocellular fat storage and improved insulin sensitivity. *Diabetes*, **59**(3), 572–579.
- Mesinovic, J., Zengin, A., De Courten, B., Ebeling, P. R., & Scott, D. (2019). Sarcopenia and type 2 diabetes mellitus: A bidirectional relationship. *Diabetes, Metabolic Syndrome and Obesity*, **12**, 1057–1072.
- Messinger, R. B., Naik, A. K., Jagodic, M. M., Nelson, M. T., Lee, W. Y., Choe, W. J., Orestes, P., Latham, J. R., Todorovic, S. M., & Jevtovic-Todorovic, V. (2009). In vivo silencing of the CaV3.2 T-type calcium channels in sensory neurons alleviates hyperalgesia in rats with streptozocin-induced diabetic neuropathy. *Pain*, **145**(1–2), 184–95.
- Miao, Y., Xie, L., Song, J., Cai, X., Yang, J., Ma, X., Chen, S., & Xie, P. (2024). Unraveling the causes of sarcopenia: Roles of junction impairment and mitochondrial dysfunction. *Physiological Reports*, **12**(1), e15917.
- Minnock, D., Annibaldi, G., Le Roux, C. W., Contarelli, S., Krause, M., Saltarelli, R., Valli, G., Stocchi, V., Barbieri, E., & De Vito, G. (2020). Effects of acute aerobic, resistance and combined exercises on 24-h glucose variability and skeletal muscle signalling responses in type 1 diabetics. *European Journal of Applied Physiology*, **120**(12), 2677–2691.
- Minnock, D., Annibaldi, G., Valli, G., Saltarelli, R., Krause, M., Barbieri, E., & De Vito, G. (2022). Altered muscle mitochondrial, inflammatory and trophic markers, and reduced exercise training adaptations in type 1 diabetes. *The Journal of Physiology*, **600**(6), 1281–1282.

- Monaco, C. M. F., Gingrich, M. A., & Hawke, T. J. (2019). Considering type 1 diabetes as a form of accelerated muscle aging. *Exercise and Sport Sciences Reviews*, **47**(2), 98–107.
- Monti, E., Sarto, F., Sartori, R., Zanchettin, G., Löfler, S., Kern, H., Narici, M. V., & Zampieri, S. (2023). C-terminal agrin fragment as a biomarker of muscle wasting and weakness: A narrative review. *Journal of Cachexia, Sarcopenia and Muscle*, **14**(2), 730–744.
- Moreno, C., d'Almeida, O. C., Gomes, L., Paiva, I., & Castelo-Branco, M. (2023). Regular physical activity moderates the adverse impact of type 2 diabetes on brain atrophy independently from HbA1c. *Frontiers in Endocrinology*, **14**, 1135358.
- Morettini, M., Palumbo, M. C., Sacchetti, M., Castiglione, F., & Mazzà, C. (2017). A system model of the effects of exercise on plasma Interleukin-6 dynamics in healthy individuals: Role of skeletal muscle and adipose tissue. *PLoS ONE*, **12**(7), e0181224.
- Mortensen, S. P., Winding, K. M., Iepsen, U. W., Munch, G. W., Marcussen, N., Hellsten, Y., Pedersen, B. K., & Baum, O. (2019). The effect of two exercise modalities on skeletal muscle capillary ultrastructure in individuals with type 2 diabetes. *Scandinavian Journal of Medicine and Science in Sports*, **29**(3), 360–368.
- Müller, T. D., Finan, B., Bloom, S. R., D'Alessio, D., Drucker, D. J., Flatt, P. R., Fritsche, A., Gribble, F., Grill, H. J., Habener, J. F., Holst, J. J., Langhans, W., Meier, J. J., Nauck, M. A., Perez-Tilve, D., Pocai, A., Reimann, F., Sandoval, D. A., Schwartz, T. W., ... Tschöp, M. H. (2019). Glucagon-like peptide 1 (GLP-1). *Molecular Metabolism*, **30**. <https://doi.org/10.1016/j.molmet.2019.09.010>
- Muramatsu, K. (2020). Diabetes mellitus-related dysfunction of the motor system. *International Journal of Molecular Sciences*, **21**(20), 7485.
- Naclerio, F., Rodríguez-Romo, G., Barriopedro-Moro, M. I., Nez, A. J., Alvar, B. A., & Triplett, A. N. T. (2011). Control of resistance training intensity by the OMNI perceived exertion scale. *Journal of Strength & Conditioning Research*, **35**, S89–S94.
- Nesti, L., Pugliese, N. R., Sciuto, P., & Natali, A. (2020). Type 2 diabetes and reduced exercise tolerance: A review of the literature through an integrated physiology approach. In *Cardiovascular Diabetology* (Vol., **19**, Issue 1). BioMed Central Ltd.
- Ng, G. Y. Q., Loh, Z. W.-L., Fann, D. Y., Mallilankaraman, K., Arumugam, T. V., & Hande, M. P. (2024). Role of Mitogen-Activated Protein (MAP) kinase pathways in metabolic diseases. *Genome Integrity*, **15**, e20230003.
- Nojima, H., Watanabe, H., Yamane, K., Kitahara, Y., Sekikawa, K., Yamamoto, H., Yokoyama, A., Inamizu, T., Asahara, T., & Kohno, N. (2008). Effect of aerobic exercise training on oxidative stress in patients with type 2 diabetes mellitus. *Metabolism: Clinical and Experimental*, **57**(2), 170–176.
- O'Connor, K., Spendiff, S., Lochmüller, H., & Horvath, R. (2023). Mitochondrial mutations can alter neuromuscular transmission in congenital myasthenic syndrome and mitochondrial disease. *International Journal of Molecular Sciences*, **24**(10), 8505.
- Olsen, D. B., Sacchetti, M., Dela, F., Ploug, T., & Saltin, B. (2005). Glucose clearance is higher in arm than leg muscle in type 2 diabetes. *The Journal of Physiology*, **565**(2), 555–562.
- Olver, T. D., McDonald, M. W., Grisé, K. N., Dey, A., Allen, M. D., Medeiros, P. J., Lacefield, J. C., Jackson, D. N., Rice, C. L., James Melling, C. W., Noble, E. G., & Shoemaker, J. K. (2014). Exercise training enhances insulin-stimulated nerve arterial vasodilation in rats with insulin-treated experimental diabetes. *American Journal of Physiology - Regulatory Integrative and Comparative Physiology*, **306**(12), R941–950.
- Ong, K. L., Stafford, L. K., McLaughlin, S. A., Boyko, E. J., Vollset, S. E., Smith, A. E., Dalton, B. E., Duprey, J., Cruz, J. A., Hagins, H., Lindstedt, P. A., Aali, A., Abate, Y. H., Abate, M. D., Abbasian, M., Abbasi-Kangevari, Z., Abbasi-Kangevari, M., Abd ElHafeez, S., Abd-Rabu, R., ... Vos, T. (2023). Global, regional, and national burden of diabetes from 1990 to 2021, with projections of prevalence to 2050: A systematic analysis for the Global Burden of Disease Study 2021. *The Lancet*, **402**(10397), 203–234.
- Orestes, P., Osuru, H. P., McIntire, W. E., Jacus, M. O., Salajegheh, R., Jagodic, M. M., Choe, W. J., Lee, J. H., Lee, S. S., Rose, K. E., Piro, N., DiGruccio, M. R., Krishnan, K., Covey, D. F., Lee, J. H., Barrett, P. Q., Jevtic-Todorovic, V., & Todorovic, S. M. (2013). Reversal of neuropathic pain in diabetes by targeting glycosylation of Cav3.2 T-type calcium channels. *Diabetes*, **62**(11), 3828–3838.
- Orlando, G., Balducci, S., Bazzucchi, I., Pugliese, G., & Sacchetti, M. (2016). Neuromuscular dysfunction in type 2 diabetes: Underlying mechanisms and effect of resistance training. In *Diabetes/Metabolism Research and Reviews*, **32**(1), 40–50.
- Orlando, G., Balducci, S., Bazzucchi, I., Pugliese, G., & Sacchetti, M. (2017a). Muscle fatigability in type 2 diabetes. *Diabetes/Metabolism Research and Reviews*, **33**(1). <https://doi.org/10.1002/dmrr.2821>
- Orlando, G., Balducci, S., Bazzucchi, I., Pugliese, G., & Sacchetti, M. (2017b). The impact of type 1 diabetes and diabetic polyneuropathy on muscle strength and fatigability. *Acta Diabetologica*, **54**(6), 543–550.
- Orlando, G., Balducci, S., Boulton, A. J. M., Degens, H., & Reeves, N. D. (2022). Neuromuscular dysfunction and exercise training in people with diabetic peripheral neuropathy: A narrative review. *Diabetes Research and Clinical Practice*, **183**, 109183.
- Ørstavik, K., Namer, B., Schmidt, R., Schmelz, M., Hilliges, M., Weidner, C., Carr, R. W., Handwerker, H., Jørum, E., & Torebjörk, H. E. (2006). Abnormal function of C-fibers in patients with diabetic neuropathy. *Journal of Neuroscience*, **26**(44), 11287–11294.
- Osseward, P. J., & Pfaff, S. L. (2019). Cell type and circuit modules in the spinal cord. *Current Opinion in Neurobiology*, **56**, 175–184.
- Ossipov, M. H., Morimura, K., & Porreca, F. (2014). Descending pain modulation and chronification of pain. *Current Opinion in Supportive and Palliative Care*, **8**(2), 143–151.

- Ozougwu, O. (2013). The pathogenesis and pathophysiology of type 1 and type 2 diabetes mellitus. *Journal of Physiology and Pathophysiology*, **4**(4), 46–57.
- Papagianni, G., Panayiotou, C., Vardas, M., Balaskas, N., Antonopoulos, C., Tachmatzidis, D., Didangelos, T., Lambadiari, V., & Kadoglou, N. P. E. (2023). The anti-inflammatory effects of aerobic exercise training in patients with type 2 diabetes: A systematic review and meta-analysis. *Cytokine*, **164**, 156157.
- Parasoglou, P., Rao, S., & Slade, J. M. (2017). Declining skeletal muscle function in diabetic peripheral neuropathy. *Clinical Therapeutics*, **39**(6), 1085–1103.
- Pedersen, B. K. (2017). Anti-inflammatory effects of exercise: Role in diabetes and cardiovascular disease. *European Journal of Clinical Investigation*, **47**(8), 600–611.
- Pedersen, B. K., & Saltin, B. (2015). Exercise as medicine - Evidence for prescribing exercise as therapy in 26 different chronic diseases. *Scandinavian Journal of Medicine and Science in Sports*, **25**(S3), 1–72.
- Peng, B., Chen, Z., Ma, L., & Dai, Y. (2015). Cerebral alterations of type 2 diabetes mellitus on MRI: A pilot study. *Neuroscience Letters*, **606**, 100–105.
- Perry, B. D., Caldwell, M. K., Brennan-Speranza, T. C., Sbaraglia, M., Jerums, G., Garnham, A., Wong, C., Levinger, P., Asrar Ul Haq, M., Hare, D. L., Price, S. R., & Levinger, I. (2016). Muscle atrophy in patients with type 2 diabetes mellitus: Roles of inflammatory pathways, physical activity and exercise. *Exercise Immunology Review*, **22**, 94–109.
- Picconi, F., Mataluni, G., Ziccardi, L., Parravano, M., Di Renzo, A., Ylli, D., Pasqualetti, P., Studer, V., Chioma, L., Marfia, G. A., & Frontoni, S. (2018). Association between early neuroretinal dysfunction and peripheral motor unit loss in patients with type 1 diabetes mellitus. *Journal of Diabetes Research*, **2018**, 9763507.
- Pinto, A., Bergouignan, A., Dempsey, P., Roschel, H., Gualano, B., & Dunstan, D. (2023). Physiology of sedentary behavior. *Physiological Reviews*, **103**(4), 2561–2622.
- Pittenger, G. L., Ray, M., Burcus, N. I., McNulty, P., Basta, B., & Vinik, A. I. (2004). Intraepidermal nerve fibers are indicators of small-fiber neuropathy in both diabetic and nondiabetic patients. *Diabetes Care*, **27**(8), 1974–1979.
- Pop-Busui, R., Boulton, A. J. M., Feldman, E. L., Bril, V., Freeman, R., Malik, R. A., Sosenko, J. M., & Ziegler, D. (2017). Diabetic neuropathy: A position statement by the American diabetes association. *Diabetes Care*, **40**(1), 136–154.
- Praet, S. F. E., Van Loon, L. J. C., & Praet, S. (2007). Optimizing the therapeutic benefits of exercise in Type 2 diabetes. *Journal of Applied Physiology*, **103**, 1113–1120.
- Pratt, J., De Vito, G., Narici, M., & Boreham, C. (2021). Neuromuscular junction aging: A role for biomarkers and exercise. *Journals of Gerontology - Series A Biological Sciences and Medical Sciences*, **76**(4), 576–585.
- Pratt, J., De Vito, G., Segurado, R., Pessanha, L., Dolan, J., Narici, M., & Boreham, C. (2022). Plasma neurofilament light levels associate with muscle mass and strength in middle-aged and older adults: Findings from GenoFit. *Journal of Cachexia, Sarcopenia and Muscle*, **13**(3), 1811–1820.
- Prior, S. J., Blumenthal, J. B., Katzel, L. I., Goldberg, A. P., & Ryan, A. S. (2014). Increased skeletal muscle capillarization after aerobic exercise training and weight loss improves insulin sensitivity in adults with IGT. *Diabetes Care*, **37**(5), 1469–1475.
- Proske, U., & Gandevia, S. C. (2012). The proprioceptive senses: Their roles in signaling body shape, body position and movement, and muscle force. *Physiological Reviews*, **92**(4), 1651–1697.
- Qaisar, R., Javed, M., Khan, I. M., Ahmad, F., & Karim, A. (2024). Metformin improves skeletal muscle and physical capacity by stabilizing neuromuscular junction in older adults. *Archives of Gerontology and Geriatrics*, **127**, 105587.
- Qi, C., Song, X., Wang, H., Yan, Y., & Liu, B. (2022). The role of exercise-induced myokines in promoting angiogenesis. *Frontiers in Physiology*, **13**, 981577.
- Racha, P., Selvam, S., Bose, B., Bantwal, G., & Sambashivaiah, S. (2022). Circulating CTerminal agrin fragment: A potential marker for sarcopenia among type 2 diabetes. *Indian Journal of Endocrinology and Metabolism*, **26**(4), 334–340.
- Ramamurthy, B., Höök, P., Jones, D., & Larsson, L. (2001). Changes in myosin structure and function in response to glycation. *The Federation of American Societies for Experimental Biology Journal*, **15**(13), 2415–2422.
- Ramamurthy, B., Jones, A. D., & Larsson, L. (2003). Glutathione reverses early effects of glycation on myosin function. *American Journal of Physiology - Cell Physiology*, **285**(2), C419–424.
- Ramji, N., Toth, C., Kennedy, J., & Zochodne, D. W. (2007). Does diabetes mellitus target motor neurons? *Neurobiology of Disease*, **26**(2), 301–311.
- Rask-Madsen, C., & King, G. L. (2013). Vascular complications of diabetes: Mechanisms of injury and protective factors. *Cell Metabolism*, **17**(1), 20–33.
- Reske-Nielsen, E., Lundbæk, K., & Rafaelsen, O. J. (1966). Pathological changes in the central and peripheral nervous system of young long-term diabetics - I. Diabetic encephalopathy. *Diabetologia*, **1**(3–4), 233–241.
- Reusch, J. E. B., Bridenstine, M., & Regensteiner, J. G. (2013). Type 2 diabetes mellitus and exercise impairment. *Reviews in Endocrine and Metabolic Disorders*, **14**(1), 77–86.
- Richter, E. A., & Hargreaves, M. (2013). Exercise, GLUT4, and skeletal muscle glucose uptake. *Physiological Reviews*, **93**, 993–1017.
- Riddell, M. C., Gallen, I. W., Smart, C. E., Taplin, C. E., Adolffson, P., Lumb, A. N., Kowalski, A., Rabasa-Lhoret, R., McCrimmon, R. J., Hume, C., Annan, F., Fournier, P. A., Graham, C., Bode, B., Galassetti, P., Jones, T. W., Millán, I. S., Heise, T., Peters, A. L., ... Laffel, L. M. (2017). Exercise management in type 1 diabetes: A consensus statement. *The Lancet Diabetes and Endocrinology*, **5**(5), 377–390.
- Robbins, J. M., & Gerszten, R. E. (2023). Exercise, exerkines, and cardiometabolic health: From individual players to a team sport. *Journal of Clinical Investigation*, **133**(11), e168121.

- Roberts, M. D., McCarthy, J. J., Hornberger, T. A., Phillips, S. M., Mackey, A. L., Nader, G. A., Boppart, M. D., Kavazis, A. N., Reidy, P. T., Ogasawara, R., Libardi, C. A., Ugrinowitsch, C., Booth, F. W., & Esser, K. A. (2023). Mechanisms of mechanical overload-induced skeletal muscle hypertrophy: Current understanding and future directions. In *Physiological reviews* (Vol., 103, Issue 4, pp. 2679–2757). NLM (Medline).
- Robertson, R. P., Harmon, J., Tran, P. O., Tanaka, Y., & Takahashi, H. (2003). Perspectives in diabetes glucose toxicity in-cells: Type 2 diabetes, good radicals gone bad, and the glutathione connection. *Perspective in Diabetes*, 52(3), 581–587.
- Rodino-Klapac, L. R., Haidet, A. M., Kota, J., Handy, C., Kaspar, B. K., & Mendell, J. R. (2009). Inhibition of myostatin with emphasis on follistatin as a therapy for muscle disease. In *Muscle and Nerve*, 39(3), 283–296.
- Röhling, M., Herder, C., Roden, M., Stemper, T., & Müssig, K. (2016). Effects of long-term exercise interventions on glycaemic control in type 1 and type 2 diabetes: A systematic review. *Experimental and Clinical Endocrinology and Diabetes*, 124(8), 487–494.
- Rossetti, L., Giaccari, A., & Defronzo, R. A. (1990). Glucose toxicity. *Diabetes Care*, 13(6), 610–630.
- Sacchetti, M. S., Balducci, S., Bazzucchi, I., Carlucci, F., Di Palumbo, A. S., Haxhi, J., Conti, F., Di Biase, N., Calandriello, E., & Pugliese, G. (2013). Neuromuscular dysfunction in diabetes: Role of nerve impairment and training status. *Medicine and Science in Sports and Exercise*, 45(1), 52–59.
- Saeedi, P., Petersohn, I., Salpea, P., Malanda, B., Karuranga, S., Unwin, N., Colagiuri, S., Guariguata, L., Motala, A. A., Ogurtsova, K., Shaw, J. E., Bright, D., & Williams, R. (2019). Global and regional diabetes prevalence estimates for 2019 and projections for 2030 and 2045: Results from the International Diabetes Federation Diabetes Atlas, 9th edition. *Diabetes Research and Clinical Practice*, 157, 107843.
- Said, G. (2007). Diabetic neuropathy - A review. *Nature Clinical Practice Neurology*, 3(6), 331–340.
- Salter, M. W., & Beggs, S. (2014). Sublime microglia: Expanding roles for the guardians of the CNS. *Cell*, 158(1), 15–24.
- Sango, K., Mizukami, H., Horie, H., & Yagihashi, S. (2017). Impaired axonal regeneration in diabetes. Perspective on the underlying mechanism from in vivo and in vitro experimental studies. *Frontiers in Endocrinology*, 8, 12.
- Schwensfeier, L., Kreutz, T., & Brinkmann, C. (2023). Is there a different rating of perceived exertion in men with type 2 diabetes mellitus? *Journal of Diabetes and Metabolic Disorders*, 22(2), 1399–1404.
- Selvarajah, D., Kar, D., Khunti, K., Davies, M. J., Scott, A. R., Walker, J., & Tesfaye, S. (2019). Diabetic peripheral neuropathy: Advances in diagnosis and strategies for screening and early intervention. *The Lancet Diabetes and Endocrinology*, 7(12), 938–948.
- Selvarajah, D., Sloan, G., Teh, K., Wilkinson, I. D., Heiberg-Gibbons, F., Awadh, M., Kelsall, A., Grieg, M., Pallai, S., & Tesfaye, S. (2023). Structural brain alterations in key somatosensory and nociceptive regions in diabetic peripheral neuropathy. *Diabetes Care*, 46(4), 777–785.
- Selvarajah, D., Wilkinson, I. D., Davies, J., Gandhi, R., & Tesfaye, S. (2011). Central nervous system involvement in diabetic neuropathy. *Current Diabetes Reports*, 11(4), 310–322.
- Selvarajah, D., Wilkinson, I. D., Emery, C. J., Harris, N. D., Shaw, P. J., Witte, D. R., Griffiths, P. D., & Tesfaye, S. (2006). Early involvement of the spinal cord in diabetic peripheral neuropathy. *Diabetes Care*, 29(12), 2664–2669.
- Selvarajah, D., Wilkinson, I. D., Maxwell, M., Davies, J., Sankar, A., Boland, E., Gandhi, R., Tracey, I., & Tesfaye, S. (2014). Magnetic resonance neuroimaging study of brain structural differences in diabetic peripheral neuropathy. *Diabetes Care*, 37(6), 1681–1688.
- Sempere-Bigorra, M., Julián-Rochina, I., & Cauli, O. (2021). Differences and similarities in neuropathy in type 1 and 2 diabetes: A systematic review. *Journal of Personalized Medicine*, 11(3), 230.
- Senefeld, J. W., Keenan, K. G., Ryan, K. S., D'Astice, S. E., Negro, F., & Hunter, S. K. (2020). Greater fatigability and motor unit discharge variability in human type 2 diabetes. *Physiological Reports*, 8(13), e14503.
- Seok, W. P., Goodpaster, B. H., Strotmeyer, E. S., Kuller, L. H., Broudeau, R., Kammerer, C., de Rekeneire, N., Harris, T. B., Schwartz, A. V., Tylavsky, F. A., Yong-Wook, C., & Newman, A. B. (2007). Accelerated loss of skeletal muscle strength in older adults with type 2 diabetes: The health, aging, and body composition study. *Diabetes Care*, 30(6), 1507–1512.
- Seung, W. L., Kyoung, E. S., Dong, S. S., Sung, M. A., Eun, S. H., Dae, J. K., Moon, S. N., & Lee, K. W. (2005). Alterations in peripheral blood levels of TIMP-1, MMP-2, and MMP-9 in patients with type-2 diabetes. *Diabetes Research and Clinical Practice*, 69(2), 175–179.
- Shankarappa, S. A., Piedras-Rentería, E. S., & Stubbs, E. B. (2011). Forced-exercise delays neuropathic pain in experimental diabetes: Effects on voltage-activated calcium channels. *Journal of Neurochemistry*, 118(2), 224–236.
- Shaw, J. E., Sicree, R. A., & Zimmet, P. Z. (2010). Global estimates of the prevalence of diabetes for 2010 and 2030. *Diabetes Research and Clinical Practice*, 87(1), 4–14.
- Sherrington, C. S. (1906). *The Integrative Action of the Nervous System*. CT, Yale University.
- Shun, C. T., Chang, Y. C., Wu, H. P., Hsieh, S. C., Lin, W. M., Lin, Y. H., Tai, T. Y., & Hsieh, S. T. (2004). Skin denervation in type 2 diabetes: Correlations with diabetic duration and functional impairments. *Brain*, 127(7), 1593–1605.
- Sima, A. A. F., & Brismar, T. (1985). Reversible diabetic nerve dysfunction: Structural correlates to electrophysiological abnormalities. *Annals of Neurology*, 18(1), 21–29.
- Singh, R., Kishore, L., & Kaur, N. (2014). Diabetic peripheral neuropathy: Current perspective and future directions. *Pharmacological Research*, 80, 21–35.
- Sjøberg, K. A., Frøsig, C., Kjøbsted, R., Sylow, L., Kleinert, M., Betik, A. C., Shaw, C. S., Kiens, B., Wojtaszewski, J. F. P., Rattigan, S., Richter, E. A., & McConell, G. K. (2017). Exercise increases human skeletal muscle insulin sensitivity via coordinated increases in microvascular perfusion and molecular signaling. *Diabetes*, 66(6), 1501–1510.

- Škarabot, J., Brownstein, C. G., Casolo, A., Del Vecchio, A., & Ansdell, P. (2021). The knowns and unknowns of neural adaptations to resistance training. In *European Journal of Applied Physiology* (Vol., **121**, Issue 3, pp. 675–685). Springer Science and Business Media Deutschland GmbH.
- Song, B. X., Azhar, L., Koo, G. K. Y., Marzolini, S., Gallagher, D., Swardfager, W., Chen, C., Ba, J., Herrmann, N., & Lanctôt, K. L. (2024). The effect of exercise on blood concentrations of angiogenesis markers in older adults: A systematic review and meta-analysis. *Neurobiology of Aging*, **135**. <https://doi.org/10.1016/j.neurobiolaging.2023.12.004>
- Streckmann, F., Balke, M., Cavaletti, G., Toscanelli, A., Bloch, W., Décard, B. F., Lehmann, H. C., & Faude, O. (2022). Exercise and neuropathy: Systematic review with meta-analysis. In *Sports Medicine* (Vol., **52**, Issue 5, pp. 1043–1065). Springer Science and Business Media Deutschland GmbH.
- Su, J.-B., Zhao, L.-H., Zhang, X.-L., Cai, H.-L., Huang, H.-Y., Xu, F., Chen, T., & Wang, X.-Q. (2018). HbA1c variability and diabetic peripheral neuropathy in type 2 diabetic patients. *Cardiovascular Diabetology*, **17**(1), 47.
- Su, W., Tao, M., Ma, L., Tang, K., Xiong, F., Dai, X., & Qin, Y. (2023). Dose-response relationships of resistance training in Type 2 diabetes mellitus: A meta-analysis of randomized controlled trials. In *Frontiers in Endocrinology* (Vol., **14**). Frontiers Media SA.
- Su, X., He, J., Cui, J., Li, H., & Men, J. (2022). The effects of aerobic exercise combined with resistance training on inflammatory factors and heart rate variability in middle-aged and elderly women with type 2 diabetes mellitus. *Annals of Noninvasive Electrocardiology*, **27**(6), e12996.
- Suzuki, A., Yabu, A., & Nakamura, H. (2022). Advanced glycation end products in musculoskeletal system and disorders. *Methods (San Diego, California)*, **203**, 179–186.
- Syeda, U. S. A., Battillo, D., Visaria, A., & Malin, S. K. (2023). The importance of exercise for glycemic control in type 2 diabetes. *American Journal of Medicine Open*, **9**, 100031.
- Tan, A. M., Samad, O. A., Fischer, T. Z., Zhao, P., Persson, A. K., & Waxman, S. G. (2012). Maladaptive dendritic spine remodeling contributes to diabetic neuropathic pain. *Journal of Neuroscience*, **32**(20), 6795–807.
- Tao, Z. Y., Wang, P. X., Wei, S. Q., Traub, R. J., Li, J. F., & Cao, D. Y. (2019). The role of descending pain modulation in chronic primary pain: Potential application of drugs targeting serotonergic system. *Neural Plasticity*, **2019**, 1389296.
- Terada, M., Yasuda, H., & Kikkawa, R. (1998). Delayed Wallerian degeneration and increased neuro-filament phosphorylation in sciatic nerves of rats with streptozocin-induced diabetes. *Journal of the Neurological Sciences*, **155**(1), 23–30.
- Treadway, J. L., James, D. E., Burcel, E., & Ruderman, N. B. (1989). Effect of exercise on insulin receptor binding and kinase activity in skeletal muscle. *American Journal of Physiology - Endocrinology and Metabolism*, **256**(1), E138–144.
- Umpierre, D., Ribeiro, P. A. B., Kramer, C. K., Leitão, C. B., Zucatti, A. T. N., Azevedo, M. J., Gross, J. L., Ribeiro, J. P., & Schaan, B. D. (2011). Physical activity advice only or structured exercise training and association with HbA1c levels in type 2 diabetes: A systematic review and meta-analysis. *Journal of the American Medical Association*, **305**(17), 1790–1799.
- Valli, G., Wu, R., Minnock, D., Sirago, G., Annibalini, G., Casolo, A., Del Vecchio, A., Toniolo, L., Barbieri, E., & de Vito, G. (2025). Can non-invasive motor unit analysis reveal distinct neural strategies of force production in young with uncomplicated type 1 diabetes? *European Journal of Applied Physiology*, **12**, 247–259.
- Van Dijk, J., & Van Loon, L. J. C. (2015). Exercise strategies to optimize glycemic control in type 2 diabetes: A continuing glucose monitoring perspective. *Diabetes Spectrum*, **28**(1), 24–31.
- Van Ryckeghem, L., Keytsman, C., De Brandt, J., Verboven, K., Verbaanderd, E., Marinus, N., Franssen, W. M. A., Frederix, I., Bakelants, E., Petit, T., Jogani, S., Stroobants, S., Dendele, P., Bito, V., Verwerf, J., & Hansen, D. (2022). Impact of continuous vs. interval training on oxygen extraction and cardiac function during exercise in type 2 diabetes mellitus. *European Journal of Applied Physiology*, **122**(4), 875–887.
- Ved, N., da Vitoria Lobo, M. E., Bestall, S. M., Vidueira, C. L., Beazley-Long, N., Ballmer-Hofer, K., Hirashima, M., Bates, D. O., Donaldson, L. F., & Hulse, R. P. (2018). Diabetes-induced microvascular complications at the level of the spinal cord: A contributing factor in diabetic neuropathic pain. *The Journal of Physiology*, **596**(16), 3675–3693.
- Watanabe, K., Miyamoto, T., Tanaka, Y., Fukuda, K., & Moritani, T. (2012). Type 2 diabetes mellitus patients manifest characteristic spatial EMG potential distribution pattern during sustained isometric contraction. *Diabetes Research and Clinical Practice*, **97**(3), 468–473.
- Wei, M., Gibbons, L. W., Mitchell, T. L., Kampert, J. B., Lee, C. D., & Blair, S. N. (1999). The association between cardio-respiratory fitness and impaired fasting glucose and type 2 diabetes mellitus in men. *Annals of Internal Medicine*, **130**(2), 89–96.
- West, S. J., Bannister, K., Dickenson, A. H., & Bennett, D. L. (2015). Circuitry and plasticity of the dorsal horn - Toward a better understanding of neuropathic pain. *Neuroscience*, **300**, 254–275.
- Wilson, G. A., Wilkins, G. T., Cotter, J. D., Lamberts, R. R., Lal, S., & Baldi, J. C. (2019). HIIT improves left ventricular exercise response in adults with type 2 diabetes. *Medicine and Science in Sports and Exercise*, **51**(6), 1099–1105.
- Wong, K. H., Chow, M. B. C. Y., Lui, T. H., Cheong, Y. K., & Tam, K. F. (2017). Denervation pseudohypertrophy of calf muscles associated with diabetic neuropathy. *Radiology Case Reports*, **12**(4), 815–820.
- Woolf, C. J. (2011). Central sensitization: Implications for the diagnosis and treatment of pain. *Pain*, **152**(Suppl 3), S2–S15.
- Xu, G., Liu, B., Sun, Y., Du, Y., Snetselaar, L. G., Hu, F. B., & Bao, W. (2018). Prevalence of diagnosed type 1 and type 2 diabetes among US adults in 2016 and 2017: Population based study. *British Medical Journal (Online)*, **362**, k1497.

- Yamaguchi, T., Kouzaki, K., Sasaki, K., & Nakazato, K. (2024). Alterations in neuromuscular junction morphology with ageing and endurance training modulate neuromuscular transmission and myofibre composition. *The Journal of Physiology*, **603**(1), 107–125.
- Yamazaki, Y., Sato, D., Yamashiro, K., Nakano, S., Onishi, H., & Maruyama, A. (2019). Acute low-intensity aerobic exercise modulates intracortical inhibitory and excitatory circuits in an exercised and a non-exercised muscle in the primary motor cortex. *Frontiers in Physiology*, **10**, 1361.
- Yang, C., Zhao, X., An, X., Zhang, Y., Sun, W., Zhang, Y., Duan, Y., Kang, X., Sun, Y., Jiang, L., & Lian, F. (2023). Axonal transport deficits in the pathogenesis of diabetic peripheral neuropathy. *Frontiers in Endocrinology*, **14**, 1136796.
- Yang, D., Yang, Y., Li, Y., & Han, R. (2019). Physical exercise as therapy for type 2 diabetes mellitus: From mechanism to orientation. *Annals of Nutrition and Metabolism*, **74**(4), 313–321.
- Yapanis, M., James, S., Craig, M. E., O'Neal, D., & Ekinici, E. I. (2022). Complications of diabetes and metrics of glycemic management derived from continuous glucose monitoring. *Journal of Clinical Endocrinology and Metabolism*, **107**(6), e2221–e2236.
- Yardley, J. E., Kenny, G. P., Perkins, B. A., Riddell, M. C., Malcolm, J., Boulay, P., Khandwala, F., & Sigal, R. J. (2012). Effects of performing resistance exercise before versus after aerobic exercise on glycemia in type 1 diabetes. *Diabetes Care*, **35**(4), 669–675.
- Yorek, M. S., Obrosova, A., Shevalye, H., Lupachyk, S., Harper, M. M., Kardon, R. H., & Yorek, M. A. (2014). Effect of glycemic control on corneal nerves and peripheral neuropathy in streptozotocin-induced diabetic C57Bl/6J mice. *Journal of the Peripheral Nervous System*, **19**(3), 205–217.
- You, W., Knoop, K., Berendschot, T. T. J. M., Benedikter, B. J., Webers, C. A. B., Reutelingsperger, C. P. M., & Gorgels, T. G. M. F. (2024). PGC-1 α mediated mitochondrial biogenesis promotes recovery and survival of neuronal cells from cellular degeneration. *Cell Death Discovery*, **10**(1), 180.
- Zaccardi, F., O'Donovan, G., Webb, D. R., Yates, T., Kurl, S., Khunti, K., Davies, M. J., & Laukkanen, J. A. (2015). Cardiorespiratory fitness and risk of type 2 diabetes mellitus: A 23-year cohort study and a meta-analysis of prospective studies. *Atherosclerosis*, **243**(1), 131–137.
- Zacker, R. (2005). Strength training in diabetes management. *Diabetes Spectrum*, **18**(2), 71.
- Zaharieva, D. P., & Addala, A. (2022). Current and novel strategies to reduce fear of hypoglycemia as a barrier to physical activity in adults and youth with type 1 diabetes. *Canadian Journal of Diabetes*, **46**(1), 1–2.
- Zang, Y., Jiang, D., Zhuang, X., & Chen, S. (2023). Changes in the central nervous system in diabetic neuropathy. *Heliyon*, **9**(8), e18368.
- Zanusso, S., Sacchetti, M., Sundberg, C. J., Orlando, G., Benvenuti, P., & Balducci, S. (2017). Exercise in type 2 diabetes: Genetic, metabolic and neuromuscular adaptations. A review of the evidence. In *British Journal of Sports Medicine* (Vol. 51, Issue 21, pp. 1533–1538). BMJ Publishing Group.
- Zhang, T., Shaw, M., & Cherbuin, N. (2022). Association between Type 2 Diabetes Mellitus and Brain Atrophy: A Meta-Analysis. *Diabetes and Metabolism Journal*, **46**(5), 781–802.
- Zhang, Y. H., Hu, H. Y., Xiong, Y. C., Peng, C., Hu, L., Kong, Y. Z., Wang, Y. L., Guo, J. B., Bi, S., Li, T. S., Ao, L. J., Wang, C. H., Bai, Y. L., Fang, L., Ma, C., Liao, L. R., Liu, H., Zhu, Y., Zhang, Z. J., ... Wang, X. Q. (2021). Exercise for Neuro-pathic Pain: A Systematic Review and Expert Consensus. *Frontiers in Medicine*, **8**, 756940.
- Zhao, X., He, Q., Zeng, Y., & Cheng, L. (2021). Effectiveness of combined exercise in people with type 2 diabetes and concurrent overweight/obesity: A systematic review and meta-analysis. *British Medical Journal Open*, **11**(10), e046252.
- Zhu, J., Hu, Z., Luo, Y., Liu, Y., Luo, W., Du, X., Luo, Z., Hu, J., & Peng, S. (2024). Diabetic peripheral neuropathy: Pathogenetic mechanisms and treatment. *Frontiers in Endocrinology*, **14**, 1265372.
- Zochodne, D. W., Verge, V. M. K., Cheng, C., Sun, H., & Johnston, J. (2001). Does diabetes target ganglion neurones? Progressive sensory neurone involvement in long-term experimental diabetes. *Brain*, **124**(11), 2319–2334.

Additional information

Competing interests

The authors declare that they have no competing interests.

Author contributions

E.L. designed and conceived this review. All authors drafted and critically revised the article. All authors have approved the final version of the manuscript submitted for publication and agree to be accountable for all aspects of the work. All persons designated as authors qualify for authorship, and all those who qualify for authorship are listed.

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Supporting information

Additional supporting information can be found online in the Supporting Information section at the end of the HTML view of the article. Supporting information files available:

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